

Recent advances in nanomedicine for ocular drug delivery

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ABSTRACT

Vision impairment is a major global health challenge, with its prevalence projected to rise significantly in the coming decades due to an aging population and increasing rates of chronic diseases. Ocular conditions such as age-related macular degeneration, cataracts, refractive errors, glaucoma, and diabetic retinopathy are among the primary causes of vision loss, collectively affecting nearly 200 million individuals worldwide. This growing burden has intensified the demand for ophthalmic therapies that are more effective, safer, and more targeted. Among existing treatment strategies, ocular drug delivery systems provide a non-invasive route for administering medications directly to ocular tissues. However, their clinical effectiveness is often compromised by various anatomical and physiological barriers, including tear turnover, blinking, nasolacrimal drainage, and blood-ocular barriers, which limit drug retention time and significantly reduce bioavailability. In response to these challenges, the application of nanomedicine has emerged as a highly promising strategy to improve ocular drug delivery. This review presents recent advances in drug nanodelivery systems – such as dendrimers, liposomes, nanoemulsion, solid lipid nanoparticles, *in situ* gel formulations, exosomes, metal-organic frameworks, and nanocrystals – that have demonstrated advantages in enhancing drug solubility, prolonging drug release, improving corneal penetration, and reducing dosing frequency and systemic side effects. In addition, the integration of artificial intelligence (AI) and personalized medicine in the development and optimization of ocular nanomedicine is explored. AI tools such as predictive modeling, machine learning algorithms, and data-driven formulation strategies remain underutilized in ophthalmology, yet they offer tremendous potential to accelerate innovation, individualize treatment, and enhance clinical translation. This review concludes that future research should prioritize not only the advancement of safer and more efficient drug nanodelivery systems but also the incorporation of AI to transform ocular drug delivery into a more precise and patient-centered approach.

Keywords:

Eye drops; nanocarrier; nanoparticle; nanotechnology; ocular drug delivery

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1. Introduction

Vision is widely regarded as the most vital sensory modality as it plays an integral role in our daily lives, making vision impairment caused by eye diseases particularly impactful and debilitating. According to the World Health Organization database, an estimated 2.2 billion people suffer from near or distance vision impairment, of which 1 billion cases could have been prevented with appropriate intervention.¹ Among these

preventable cases, cataracts account for 94 million, uncorrected refractive errors for 88 million, and age-related macular degeneration, glaucoma, and diabetic retinopathy for 8 million, 7 million, and 4 million cases, respectively. Globally, the prevalence of ocular diseases has been observed to increase as the population grows. In addition, vision impairment has reportedly resulted in an economic burden of US\$411 billion.² As such, the need for solutions to combat vision loss has gained increasing traction.

Patients have preferred topical delivery of ocular drugs as the drug administration method over others, such as intraocular delivery and intravitreal injection. This preference could be attributed to the ease of non-invasive administration and better patient adherence. Nevertheless, the distinctive structure and function of the eye significantly limit the absorption of ocular medications, with bioavailability often being below 5%. The primary obstacles hindering progress in drug delivery are the static ocular barriers located within the posterior and anterior segments of the eye (**Figure 1**).³ Developing effective strategies to bypass these barriers has been a longstanding challenge for researchers.

In response to these challenges, drug nanodelivery systems have emerged as a novel drug delivery method. Nanodelivery systems offer several advantages, such as prolonged ocular residence times, increased corneal permeability, and sustained drug release, making them an attractive drug delivery modality. This review explores various drug nanodelivery systems, such as microemulsions, solid lipid nanoparticles, nanoemulsions, liposomes, *in situ* gels, dendrimers, exosomes, metal-organic frameworks (MOF), and nanocrystals, designed to overcome the challenges caused by the eye's anatomy and physiology (**Figure 2**).

2. Barriers to ocular delivery

The human eye functions as a sensory organ responsible for detecting and processing visual input. The eye consists primarily of two regions, the posterior and anterior segments. The cornea, conjunctiva, ciliary body, aqueous humor, and lens collectively form the anterior segment of the eye. The posterior segment consists of the retina, sclera, and choroid. Following topical drug administration, the majority of the active ingredients are eliminated by the precorneal tear film.⁴ The tear film consists of a lipid layer and an aqueous mucous layer; the amphiphilic properties of the tear film hinder the penetration of purely hydrophilic and hydrophobic substances. Despite being the most widely utilized method for ocular drug delivery, topical administration suffers from low bioavailability (1 – 5%) in the anterior segment due to the combined effects of tear film clearance and eyelid blinking. These physiological factors not only limit drug absorption but may also necessitate frequent dosing to maintain therapeutic levels. Furthermore, nasolacrimal drainage into the systemic circulation after instillation could also result in decreased bioavailability and undesired side effects.⁵ The administration process may induce irritation or discomfort, triggering reflex tear flow that further decreases drug retention and negatively impacts patient compliance.

The three principal layers of the cornea include the epithelium, stroma, and endothelium. The cornea is a negatively charged membrane at physiological pH; thus, positively charged molecules can penetrate the cornea more easily than negatively

charged molecules. The corneal epithelium is composed of basal cells, wing cells, and squamous cells, all of which are interconnected by tight junctions that restrict the diffusion of larger molecules. The endothelium is notably hydrophobic and is the most significant barrier to drug penetration. Positioned between the aqueous humor and the stroma, the corneal endothelium facilitates the transport of macromolecules between these layers. The endothelium, like the epithelium, is hydrophobic and consists of tightly packed cells, albeit it is formed by a monolayer of flattened epithelial-like cells. The epithelium and endothelium form the blood-aqueous barrier, which serves to limit the selective diffusion of different solutes through the neighboring cells.⁶ In contrast to the corneal epithelium and endothelium, the stroma is hydrophilic and is formed by tightly packed collagen. The hydrophilic nature of the stroma allows it to serve as a barrier, preventing hydrophobic molecules from passing deeper into the eye. This biphasic environment indicates that the cornea requires ocular formulations with amphipathic properties and dual-phase solubility for ocular administration.

The conjunctiva is another part of the anterior segment. It is a transparent and thin membrane, which has been noted to be more permeable to drugs than the cornea. Hydrophilic drugs are more permeable to the conjunctiva, though it is uncertain whether hydrophobic drugs are less permeable. The ciliary body, composed of smooth muscle, serves two key roles: The secretion of aqueous humor – which delivers nutrients to avascular tissues and regulates intraocular pressure – and waste drainage from the cornea and lens, including ocular drugs. Drug elimination in the anterior segment is facilitated by the aqueous humor turnover, which is secreted by the ciliary body.

Similar to the anterior segment, drug delivery to the posterior segment of the eye is challenging due to the presence of multiple barriers. The sclera, the white outer layer of the eye, is a long tissue located slightly below the conjunctiva. It is primarily made up of an extracellular matrix consisting of collagen fibrils and glycoproteins.⁷ Compared to the cornea, the sclera permits greater solute diffusion, mainly through transscleral diffusion, allowing larger molecules to traverse the porous spaces within the collagen structure, which ranges in diameter from 25 to 300 nm. It is important to note that transscleral permeability is significantly affected by molecular charge, with negatively charged molecules exhibiting higher permeability through the sclera compared to positively charged ones. The choroid, found between the sclera and retina, contains a dense network of capillaries and is reinforced by the Bruch's membrane. It is a vascular tissue that mainly supplies nutrients to the retina. The Bruch's membrane-choroid complex acts as a stronger barrier to drug delivery than the sclera through the transscleral route. It is also more selective than the sclera, as solutions tend to bind to the tissue, thereby reducing overall drug efficiency.⁸

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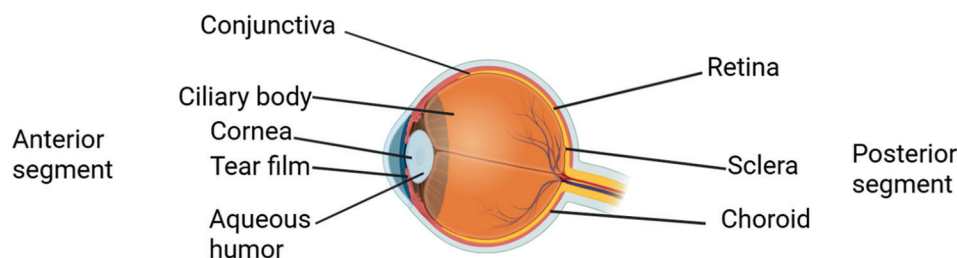


Figure 1. Anatomical barriers to ocular drug delivery. The illustration categorizes key components of the anterior and posterior eye segments that hinder effective drug transport, such as the tear film, cornea, and retina. Understanding these physiological barriers is crucial for designing efficient ocular delivery strategies. Artwork created with BioRender (<https://BioRender.com/n50y555>).

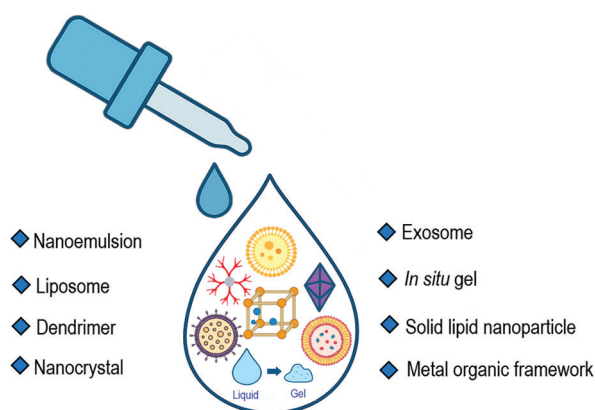


Figure 2. Overview of drug nanodelivery systems explored in this review for ocular drug delivery, each offering specific advantages in enhancing ocular drug bioavailability and retention. Artwork generated with ChatGPT and external design tools.

The retina is a transparent, thin layer of photoreceptor cells situated in the innermost region of the eye. It consists of the inner neural retina and the retinal pigment epithelium. The retina has been observed to progressively hinder the entry of larger molecules due to the multiple layers that form the retinal pigment epithelium and inner neural retina.⁹ **Table 1** provides a summary of the major ocular barriers in the anterior and posterior segments that restrict drug penetration and bioavailability.

3. Types of nanodelivery systems used in ocular therapy

3.1. Dendrimers

Dendrimers are a class of star-shaped, nano-sized polymers with a branching web-like structure. They possess a terminal end group on each branch that can be modified for functionalization. They are mainly used to increase the specificity of compounds by altering the pharmacokinetic and pharmacodynamic properties of a drug.¹⁰ For ocular delivery, specifically, it is effective when incorporated into hydrogels and when combined with polyethylene glycol groups, using it primarily for targeted drug delivery.¹¹ **Figure 3** shows the most used structural basis for dendrimers, which is the poly(amidoamine) structure. The most common synthesis methods for dendrimers are categorized into convergent, divergent, or click chemistry approaches. For the divergent method, several monomeric modules are assembled and added

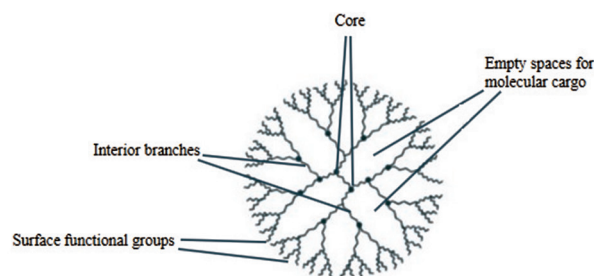


Figure 3. General architecture of a poly(amidoamine) dendrimer, highlighting its branched structure and drug-encapsulating interior. Artwork created with BioRender (<https://BioRender.com/n50y555>).

Table 1. Ocular barriers of the eye

| Segments of the eye | Description |
|------------------------------|--|
| Posterior segment of the eye | |
| Retina | Hinders the entry of large molecules due to multiple layers |
| Sclera | Permeability is predicated on the charge of the molecule |
| Choroid | Part of the Bruch's membrane-choroid complex, which acts as an entry barrier |
| Anterior segment of the eye | |
| Cornea | Corneal epithelium hampers large molecules. Corneal endothelium is similar to epithelium and forms the blood-aqueous barrier |
| Stroma | Hinders the entry of hydrophobic molecules |
| Conjunctiva | Seemingly more permeable to hydrophilic drugs |
| Ciliary body | Secretes aqueous humor and drains drugs from the cornea and lens |
| Aqueous humor | Aqueous humor turnover eliminates drugs |
| Tear film | Amphiphilic properties of the tear film hinder the penetration of ocular drugs |

to a core site, allowing it to grow and build outward in a branch-upon-branch structure according to certain dendritic rules and principles. However, there are several drawbacks to this method, such as side or incomplete reactions, resulting in structural defects.¹² To overcome these drawbacks, the convergent method was created, whereby several dendrons are reacted with a multi-functional core to form a dendrimer. Although the convergent method mitigated the issue regarding structural defects, it also possesses several challenges, including limited reactivity between the dendrons and the molecular nucleus due to distance, and decreased reactivity of the central

dendrons.¹³ Click chemistry involves the use of copper to join azides and alkyne to synthesize well-defined dendrimers with excellent purity and high yield.¹⁴

The treatment of diabetic retinopathy often involves invasive procedures, such as intravitreal implants delivering dexamethasone into the eye or intravitreal injections of anti-vascular endothelial growth factor agents. These approaches carry risks of ocular damage, and multiple intravitreal injections may reduce patient compliance. To overcome this issue, Alshammari *et al.*¹⁵ used poly(amidoamine) (PAMAM) dendrimer as a carrier and observed its effects on the ocular bioavailability of the investigational drug ruboxistaurin – a novel drug developed to combat diabetic retinopathy by acting as a protein kinase C beta inhibitor and inhibiting vascular endothelial growth factor release. Invasive drug delivery systems, such as direct injections to the eye, were avoided in this study in favor of non-invasive PAMAM dendrimer nanoparticles, which were designed to improve patient compliance. The PAMAM dendrimer nanoparticles demonstrated favorable drug release profiles and improved stability, and were also theorized to improve patient adherence by reducing the complications associated with invasive delivery methods.

Wang *et al.*¹⁶ formulated novel dendrimer hydrogel particles as a drug delivery carrier for anti-glaucoma drugs, brimonidine tartrate, and timolol maleate. The gel particles in the study presented several advantages, such as low toxicity, the ability to overcome the drug barriers of the eye, and minimal ocular irritation. Compared to plain drug solutions, the nanostructured dendrimer hydrogel particles demonstrated stronger corneal permeation and a greater decrease in intraocular pressure. Among the three dendrimer hydrogel particles that were tested (one nanostructured dendrimer hydrogel particle formulation and two micronized dendrimer hydrogel particles), the nanostructured dendrimer hydrogel particle demonstrated stronger efficacy. Hence, the study suggests its potential use as a carrier for other drugs.

3.2. Liposomes

Liposomes are artificial vesicles that consist of one or more phospholipid bilayers that enclose an aqueous core. Based on its size, it can be categorized into small unilamellar vesicles (10 nm – 100 nm), large unilamellar vesicles (100 nm – 1 μ m), and giant unilamellar vesicles (1 μ m or above). Unilamellar vesicles have one lipid bilayer surrounding their aqueous core, whereas multilamellar vesicles have one or more lipid bilayers,¹⁷ as displayed in **Figure 4**.

Liposomes have the capacity to encapsulate both hydrophilic and hydrophobic drugs, enabling their cellular uptake through endocytosis. Their dual ability to carry these substances makes them highly suitable for ocular drug delivery systems. Liposomes have demonstrated great efficacy in their ocular delivery to the posterior and anterior segments of the eye. In ocular drug delivery, liposomes offer additional benefits, including extended drug retention and minimal toxicity. However, a notable drawback lies in their low bioadhesiveness, which affects their ocular permeation. This hurdle can be

overcome via the integration of bioadhesive polymers such as chitosan.^{18,19}

Lai *et al.*²⁰ addressed the low stability of chrysophanol and berberine hydrochloride, compounds that have been recognized for their potential use in treating age-related macular degeneration.^{21,22} A combination of liposomes and polyamidoamine dendrimer (polyamidoamine 3.0) was used as a carrier for both these drugs. The results demonstrated an improvement in the bioavailability of berberine hydrochloride over chrysophanol–berberine hydrochloride suspension. In addition, polyamidoamine-coated liposomes were found to protect against photooxidative stress. However, their effectiveness against age-related macular degeneration requires further investigation using models specifically designed for this condition.

3.3. Nanoemulsions

Nanoemulsions are heterogeneous dispersions of two immiscible liquids and are composed of water, oil, surfactants, and co-surfactants either as oil droplets in water (o/w) nanoemulsion or water droplets in oil (w/o) nanoemulsion (**Figure 5**). Nanoemulsions serve as colloidal drug carriers, with droplet sizes typically ranging between 100 and 500 nm. Nanoemulsions differ from microemulsions in their preparation method. The preparation of nanoemulsions involves the use of thermal and/or mechanical energy, making them less thermodynamically stable. To improve stability, nanoemulsions are often paired with co-surfactants.²³ Nanoemulsions are preferred over their micro counterparts due to enhanced bioavailability, longer drug residence time, and smaller droplet size, leading to better corneal penetration.

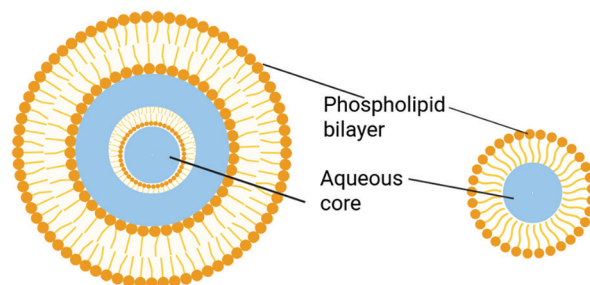


Figure 4. Comparison between unilamellar and multilamellar liposomes showing variations in bilayer configuration. Artwork created with BioRender (<https://BioRender.com/n50y555>).

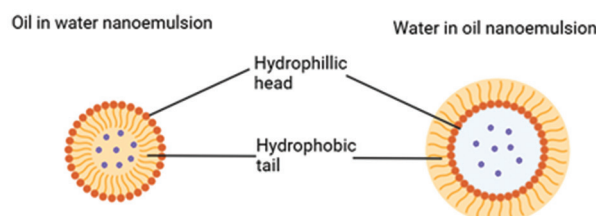


Figure 5. Visualization of oil in water and water in oil nanoemulsion types. The emulsions are stabilized by surfactant molecules that orient at the oil-water interface, with the system type determined by the dispersed phase. Artwork created with BioRender (<https://BioRender.com/n50y555>).

Nanoemulsions can be prepared using two approaches: High-energy and low-energy methods. The high-energy method involves mixing the oil, water, and surfactant for a sufficient amount of time; the resulting macroemulsion then undergoes homogenization until a suitable droplet size is achieved.^{24,25} An example of a low-energy method is the phase inversion temperature method, whereby changes in the temperature cause changes in the spontaneous curvatures of the surfactant. An o/w emulsion undergoes a phase transition as the temperature rises and water is added, resulting in the formation of water droplets dispersed within the oil phase.

Nanoemulsions are commonly used in ocular drug delivery to improve the retention and bioavailability of hydrophobic or poorly retained drugs. Moxifloxacin, a drug noted for its use in the treatment of bacterial conjunctivitis, is limited by its short residence time in the eye.²⁶ Youssef *et al.*²⁷ developed a moxifloxacin-loaded nanoemulsion formulation as well as a mucoadhesive variant and compared them to the commercially available solution. The formulations demonstrated improved permeability of moxifloxacin and increased drug residence time. The mucoadhesive variant was suggested to increase patient compliance by reducing the frequency of dosing.

3.4. Cationic nanoemulsions

Cationic nanoemulsions are biphasic formulations characterized by positively charged nanodroplets in a continuous phase. For example, in an o/w nanoemulsion, positively charged oil droplets are dispersed within the aqueous phase.²⁸ To maximize electrostatic attraction, cationic nanoemulsions are formulated with positively charged nanodroplets that interact with the negatively charged ocular mucosa. These interactions have been found to prolong drug retention in the eye, thereby enhancing therapeutic effectiveness.²⁹ Concerns regarding cationic nanoemulsions include possible irritation and toxicity due to overuse of the cationic charge inducer.^{30,31}

To address the solubility limitations of non-steroidal anti-inflammatory drugs in the treatment of dry eye disease, Jurišić Dukovski *et al.*³² developed an ibuprofen-loaded cationic nanoemulsion. The cationic o/w nanoemulsion was proven to have sufficient stability, improved drug residence time, and good biocompatibility. Another aim of the study was to stabilize the tear film layer, which is often compromised in dry eye disease.^{33,34} This was achieved through the destabilization of the nanoemulsion on encountering the tear film. The oil droplets in the nanoemulsion merged with the tear film lipid layer, while the surfactants integrated into the mucous layer, thereby restoring the tear film's integrity.

Rifampicin is one of the most potent treatments for ocular tuberculosis; however, its clinical utility is hindered by poor solubility in aqueous media.³⁵ To address this limitation, Bazán Henostroza *et al.*³⁶ investigated a cationic nanoemulsion loaded with rifampicin. The cationic rifampicin nanoemulsion demonstrated improved drug bioavailability, preserved antimicrobial properties, and enhanced patient quality of life due to reduced frequency of product instillation.

3.5. Solid lipid nanocarriers

Solid lipid nanocarriers are classified into two types: Solid lipid nanoparticles and nanostructured lipid carriers. The key distinction between them is their composition at room temperature. Solid lipid nanocarriers consist of lipids that remain solid, while nanostructured lipid carriers contain a combination of solid and liquid lipids.³⁷

Solid lipid nanocarriers are commonly prepared using high-pressure homogenization, solvent emulsification-evaporation, or microemulsion techniques. The high-pressure homogenization method involves using high pressure to push a liquid through a narrow gap, exposing the fluid to shear stress and cavitation forces, causing the particles to break down. The solvent emulsification-evaporation method entails dissolving soluble lipids in a water-immiscible organic solvent, followed by emulsification in an aqueous phase using high-pressure homogenization. Following homogenization, the solvent is evaporated through stirring at room temperature, resulting in the formation of lipid nanoparticles. The microemulsion method involves combining a low-melting-point fatty acid, an emulsifier, and water at a temperature above the melting point of the fatty acid. The mixture is then placed in cold water and continuously stirred.³⁸

3.5.1. Solid lipid nanoparticles

Solid lipid nanoparticles are composed of a solid lipid matrix dispersed in an aqueous medium, stabilized by a surfactant layer. Their formulation typically includes biocompatible solid lipids such as fatty acids, fatty alcohols, glycerol esters, and waxes.³⁹ The layer of surfactants helps stabilize the formulation by reducing interfacial energy between the lipid and aqueous phases during the preparation of solid lipid nanoparticles. Key benefits include low toxicity due to the use of safe excipients, a low production cost, and ease of large-scale production.⁴⁰

Khames *et al.*⁴¹ tested natamycin-loaded solid lipid nanoparticles to address the poor corneal penetration of natamycin, which is used to combat keratitis. The findings indicated that natamycin-loaded solid lipid nanoparticles exhibited enhanced efficacy compared to conventional natamycin administration, demonstrating extended drug release and superior antifungal activity against the primary fungal pathogens responsible for keratitis.^{42,43}

3.5.2. Nanostructured lipid carriers

Nanostructured lipid carriers were developed to overcome the instability issues associated with solid lipid nanoparticles. Nanostructured lipid carriers exhibit a higher loading capacity and stability, as they impede the recrystallization of solid lipids, thus preventing drug expulsion during storage.⁴⁴ Notably, nanostructured lipid carriers have been predominantly utilized for the delivery of antifungal agents.⁴⁵

Lactoferrin, a protein found in the immune system, is known to possess antifungal effects and stimulate corneal wound healing via the activation of toll-like receptors.⁴⁶ Varela-Fernández *et al.*⁴⁷ synthesized lactoferrin-loaded nanostructured lipid carriers and determined their efficacy in keratoconus

treatment. The resulting formulation exhibited low toxicity, sustained release of lactoferrin compared to a lactoferrin-buffered solution, and high stability for up to 3 months.

3.6. *In situ* gelling system

In situ gelling systems are liquid formulations that release encapsulated drugs upon undergoing a solution-to-gel transition when applied to a specific site, such as the eye, in ocular drug delivery.⁴⁸ The solution-gel phase transition is based on physiological conditions such as changes in temperature, the introduction of ions, or changes in pH.^{49,50} It functions similarly to photocages, which also release encapsulated drugs in response to stimuli (a specific wavelength of light).⁵¹

3.6.1. Thermosensitive *in situ* gel

Thermosensitive *in situ* gelling systems are typically single-phase, solution-like systems in an aqueous medium that incorporate thermosensitive polymers containing both hydrophobic and hydrophilic segments.^{52,53} These systems undergo a phase transition in response to changes in temperature. When exposed to temperatures above the lower critical solution temperature, the balance between the hydrophobic and hydrophilic parts is disrupted, leading to polymer-polymer interactions and resulting in the solution-gel phase transition (Figure 6).^{54–56}

Mahboobian *et al.*⁵⁶ investigated the use of polymers Carbopol 934 and Pluronic F127, combined with hydroxypropyl methylcellulose to create a thermosensitive *in situ* nano gel for flurbiprofen, a notably insoluble non-steroidal anti-inflammatory agent.⁵⁷ Carbopol 934 and Pluronic F127 were reported to enhance gel strength and solubility, respectively.^{58–60} The flurbiprofen nanosuspension, prepared using hydroxypropyl methylcellulose, was spray-dried to obtain a dry powder formulation designed to enhance solubility. The resulting *in situ* nano gel formulation demonstrated increased drug and corneal residence times for the flurbiprofen nanosuspension and showed good gelation at physiological temperatures.

Wang *et al.*⁶¹ synthesized a combination of thermosensitive *in situ* gel using carbon dots. Carbon dots have recently garnered traction in the medical field due to their good biocompatibility, low toxicity, and solubility in water.⁶² Diclofenac sodium, a non-steroidal anti-inflammatory drug characterized by low

ocular bioavailability, was incorporated into the system.⁶³ The drug delivery system showed potential by minimizing tear elimination, prolonging drug release, and enhancing the bioavailability of diclofenac sodium.

3.6.2. Ion-sensitive *in situ* gel

Ion-responsive polymers in ion-sensitive *in situ* gelling systems typically contain ionizable groups.^{64,65} These polymers undergo cross-linking with monovalent or divalent cations present in lacrimal fluid, where electrostatic interactions between anionic polymer chains and cations induce a solution-to-gel transformation.⁶⁶ The resulting viscosity of the gel is influenced by the concentration of available cations.⁶⁷

Luteolin, a natural flavonoid, has been investigated for its potential benefits in eye health, particularly in managing dry eye disorder.⁶⁸ Omran *et al.*⁶⁹ developed a carrageenan-based ion-sensitive *in situ* gel by incorporating oleophytocubosomes. Oleophytocubosomes are small, negatively charged particles with high entrapment efficiency⁷⁰ for the delivery of luteolin. The ion-sensitive *in situ* gel combined with oleophytocubosomes showed sustained drug release, enhanced anti-glaucoma effects, and improved anti-inflammatory effects.

3.6.3. pH-sensitive *in situ* gel

The gelation behavior of pH-sensitive *in situ* gel is determined by the measure of acidity, pKa, of the polymer.^{71,72} The pH-sensitive polymers are usually either weakly acidic or weakly basic. The solution-gel transition occurs when the pH falls below the pKa value of the weakly acidic polymer and vice versa for the weakly basic polymer.^{73,74}

As glaucoma remains a persistent clinical challenge, one of the primary treatment strategies involves reducing intraocular pressure.^{75,76} The poor bioavailability of the anti-glaucoma drug, betaxolol hydrochloride,^{77,78} in commercial eye drops has prompted the search for more effective ways to administer betaxolol hydrochloride to the eye. Allam *et al.*⁷⁹ used niosomes to encapsulate betaxolol hydrochloride combined with a pH-sensitive *in situ* gel. Niosomes are a type of colloidal delivery system similar to liposomes.^{80,81} The resulting formulation demonstrated better drug release compared to niosomes or the pH-sensitive *in situ* gel alone. Additionally, the bioavailability and residence time of betaxolol hydrochloride were increased by the formulation when compared to the commercially available eye drops. This innovative delivery system shows promise and may be further explored for its potential in managing glaucoma.

4. Other advancements

Exosomes are double-membrane vesicles secreted by cells that carry molecules such as DNA to target cells. Exosomes can also be engineered and modified to transport drugs to targeted cells. They have been noted to possess several desirable qualities in terms of ocular drug delivery, such as low cytotoxicity, high drug residence time, good targeting capacity, high drug loading capacity, and low immunogenicity.⁸² A study conducted by Cao *et al.*⁸³ utilized a microvascular endothelial cell model to study the therapeutic effects of using exosomes derived

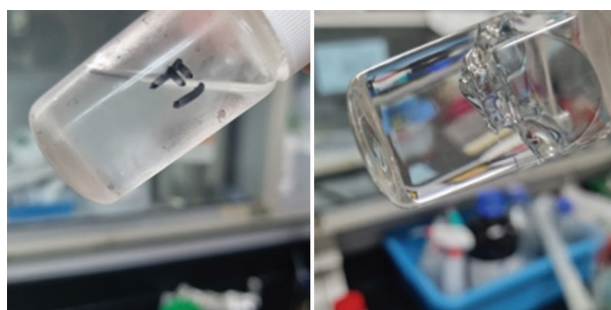


Figure 6. Visual demonstration of a temperature-responsive *in situ* gel system before (left) and after gelation (right). Upon exposure to physiological temperatures, the solution transitions into a gel phase, which prolongs ocular residence and enables sustained drug release.

from mesenchymal stem cells for the treatment of diabetic retinopathy. In this study, endothelial-mesenchymal transition and tube formation, processes associated with diabetic retinopathy, were identified through specific molecular markers. It was found that exosomes were able to suppress both endothelial-mesenchymal transition and tube formation by interacting with the microRNA-34a-5p/X-box binding protein 1 signaling pathway, indicating a potential use in the treatment of diabetic retinopathy.

Nanocrystals are nanoparticles that comprise 100% of the crystalline drug after undergoing either the top-down or bottom-up method. A simplified explanation is illustrated in **Figure 7**. The top-down method is based on the reduction in particle size, whereas the bottom-up method is based on building up molecules into small particles. Nanocrystals have been proposed for potential use in ocular applications due to their nanoscale dimensions, which contribute to improved permeability and drug bioavailability.

Nanocrystal-based ocular formulations are a relatively novel concept and have not been fully explored. Kalam *et al.*⁸⁴ investigated the potential of nanocrystals for tetragonal zirconia polycrystals, a novel 5-hydroxymethyl-oxazolidinone antibiotic. Tetragonal zirconia polycrystals-nanocrystals were found to have higher solubility in stimulated tear fluid, which indicated superior permeation and bioavailability of tetragonal zirconia polycrystals.

MOFs consist of organic linkers and interconnected metal ions to form a porous structure (**Figure 8**). Nano-sized MOFs possess a high drug loading capacity due to their porous structure. Synthetic modifications of MOFs can enhance their

targeting capacity and stability.⁸⁵ Gupta *et al.*⁸⁶ investigated the potential of MOFs for ocular drug delivery by testing a mucoadhesive MOF loaded with timolol maleate for the treatment of glaucoma. The study's results demonstrated sustained release of timolol maleate, as well as improved drug bioavailability and easy degradation in the eye, resulting in minimal side effects. These results indicate a potential use for MOFs as a drug carrier for other ocular drugs.

5. Role of artificial intelligence (AI)

In recent years, AI and machine learning have emerged as new tools to potentially facilitate the development of ocular drug nanodelivery systems. At a time when the development of novel drug formulations has been observed to be too time-consuming, expensive, and unpredictable, computational pharmaceutics has emerged as a promising approach to reduce this burden by utilizing *in silico* modeling and simulation. The use of AI and machine learning for the development of drug formulations offers several advantages, such as low costs and the elimination of ethical concerns associated with animal testing.

He *et al.*⁸⁷ collected data on the size and polydispersity index of drug nanocrystals to construct prediction models using various machine learning algorithms, such as deep neural networks, decision trees, and light gradient boosting machine. The machine learning algorithms were trained on three different preparation methods (ball wet milling, high-pressure homogenization, and antisolvent precipitation). The data used for training was split into three subsets: 80% of the data was used to construct the models, 10% was used for tuning the

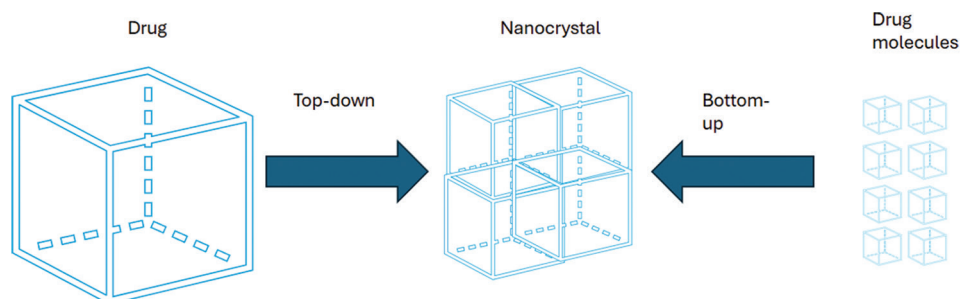


Figure 7. Schematic illustration of two synthetic routes to generate drug nanocrystals: size reduction (top-down) and molecular assembly (bottom-up). Both techniques aim to improve solubility and drug delivery performance in ocular formulations.

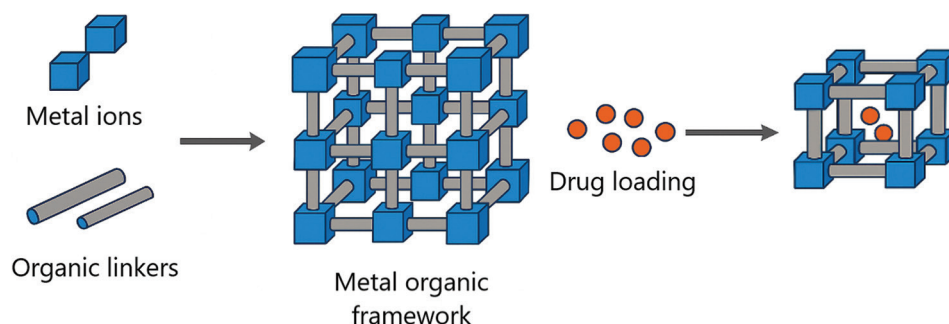


Figure 8. Conceptual workflow of metal-organic framework synthesis and post-synthetic drug loading. Their tunable porosity, high surface area, and modifiability support their use as advanced ocular drug carriers. Artwork created with ChatGPT and design tools.

model hyperparameters to ensure accuracy, and the remaining data was used as a test set for the constructed model against unfamiliar data. The results showed that the light gradient boosting machine displayed the best accuracy among all the other models. The model's efficacy in predicting the outcomes of the high-pressure homogenization method indicates a potential application for accurately forecasting the size and polydispersity index of ocular nanocarriers, including solid lipid nanoparticles, utilizing this method.

German *et al.*⁸⁸ utilized a preexisting quasi-three-dimensional model, which had individual eye segments separated into their own domains. The model was able to simulate the topical administration of eyedrops, pharmacokinetics, the physiology of aqueous humor flow, and intraocular pressure levels. The study built upon the quasi-three-dimensional model by constructing the anterior segment of the eye, including the conjunctiva, cornea, aqueous humor, ciliary body, and the lens, and simulating drug loss due to tear film turnover. The model was developed using *in vivo* studies in a rabbit eye model to observe the temporal distribution of timolol in the anterior segment. After validating the model for the rabbit's eye, they altered it to simulate the eye of a human. The predicted distribution over the course of 400 min was similar to the rabbit. The intraocular pressure and its reduction via application of timolol were simulated using an equation from a prior study conducted by a separate team, and the simulated results were compared to the actual results from said study. The intraocular pressure results demonstrated a similar pattern over 400 min to the one conducted by the separate team, with a slight under-prediction, which could be attributed to the remaining timolol concentration in the aqueous humor, preventing the intraocular pressure from returning to baseline. Although the results were satisfactory, the author noted the limitations of the model, as other factors that impact the pharmacokinetics of the eye, such as blinking, were not implemented. Regardless, the model demonstrates potential future use in ocular drug nanodelivery as a more ethical alternative to the use of animal models to study drug residence times.

6. Potential in personalized medicine

Personalized medicine, also known as precision medicine, considers a patient's unique information, such as their genetic profile, lifestyle data, current medical condition, and environmental exposure, to tailor a personalized treatment strategy.^{89,90} In ophthalmology, recent studies have utilized printing technologies to construct ocular inserts containing the nanodrug for ocular administration or to use nanoparticles as bioinks for printing onto contact lenses.

Tetyczka *et al.*⁹¹ utilized inkjet printing of itraconazole nanocrystals onto commercially available soft hydrogel contact lenses. Nanocrystals were determined to be ideal, given their good mucoadhesion to the membranes of the eye and the implications of a dual drug release profile in previous studies. Inkjet printing enables personalized dosages by delivering picoliter-scale droplets with high accuracy. The optimal formulation was compared to bulk itraconazole and demonstrated superior performance, attributed to the

nanoscale reduction of itraconazole, which enhanced drug solubility and improved inkjet printability. In addition, a dual drug release was observed after 8 h. Notably, visual clarity was preserved by intentionally excluding ink deposition in the central zone of the lens, protecting the pupil.

7. Conclusions

Despite the numerous hurdles that ocular barriers pose for drug administration, the various nanodelivery systems highlighted in this review demonstrate their efficiency and potential for administering drugs to the eye. Although significant strides have been made in nano-based topical drug delivery, future research should focus on improving formulation stability, sustaining drug release, enhancing bioavailability, and ensuring non-toxicity of the mentioned ocular drug delivery systems. Further studies on machine learning are necessary to improve the efficacy of *in silico* modeling, as it has been demonstrated to be a promising step forward for ophthalmology.

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Conflicts of interest statement

The authors declare no conflict of interest.

Author contributions

Conceptualization: NPHT and LCW; *Writing—original draft:* NPHT and LCW; *Writing—review & editing:* WML, YKL, CJL, YYT, and KWC. All authors have read and agreed to the published version of the manuscript.

Ethics approval and consent to participate

Not applicable.

Consent for publication

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Availability of data

Not applicable.

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References

1. World Health Organization. Increasing Eye Care Interventions to Address Vision Impairment: Technical Brief. *World Health Organization*; 2023.
2. Marques AP, Ramke J, Cairns J, *et al.* The economics of vision impairment and its leading causes: A systematic review. *EClinicalMedicine*. 2022;46:101354. doi: 10.1016/j.eclinm.2022.101354
3. Suri R, Beg S, Kohli K. Target strategies for drug delivery bypassing ocular barriers. *J Drug Deliv Sci Technol*. 2019;55:101389. doi: 10.1016/j.jddst.2019.101389
4. Çaprak BE, Shahbazi F, Öztürk N. Smart ocular drug delivery systems: Design principles and recent advances. *Hacettepe Univ J Faculty Pharm*. 2025;45(2):162-174. doi: 10.52794/hujpharm.1636945
5. Vaneev A, Kost O, Chesnokova N, *et al.* Nanotechnology for topical drug delivery to the anterior segment of the eye. *Int J Mol Sci*. 2021;22(22):12368. doi: 10.3390/ijms222212368
6. Dhyani A, Kumar G. A new vision to eye: Novel ocular drug delivery

- system. *Pharmacophore*. 2019;10(1):13-20. doi: 10.51847/68ngqce
7. Löscher M, Hurst J, Seiz C, Schnichels S. Topical drug delivery to the posterior segment of the eye. *Pharmaceutics*. 2022;14(1):134. doi: 10.3390/pharmaceutics14010134
 8. Lin X, Wu X, Chen X, Wang B, Xu W. Intellectual and stimulative responsive drug delivery systems in eyes. *Int J Pharm*. 2021;602:120591. doi: 10.1016/j.ijpharm.2021.120591
 9. Gote V, Sikder S, Sicotte J, Pal D. Ocular drug delivery: Present innovations and future challenges. *J Pharmacol Exp Ther*. 2019;370(3):602-624. doi: 10.1124/jpet.119.256933
 10. Mittal P, Saharan A, Verma R, et al. Dendrimers: A new race of pharmaceutical nanocarriers. *Biomed Res Int*. 2021;2021(2):8844030. doi: 10.1155/2021/8844030
 11. Wang J, Li B, Huang D, et al. Nano-in-nano dendrimer gel particles for efficient topical delivery of antiglaucoma drugs into the eye. *Chem Eng J*. 2021;425:130498. doi: 10.1016/j.cej.2021.130498
 12. Wang J, Qiao X, Li B, Yang H, Qiu L. Dendrimer-based drug delivery systems: History, challenges, and latest developments. *J Biol Eng*. 2022;16(1):18. doi: 10.1186/s13036-022-00298-5
 13. Semeraro F, Morescalchi F, Cancarini A, Russo A, Rezzola S, Costagliola C. Diabetic retinopathy, a vascular and inflammatory disease: Therapeutic implications. *Diabetes Metab*. 2019;45(6):517-527. doi: 10.1016/j.diabet.2019.04.002
 14. Najafi F, Roghani-Mamaqani H, Salami-Kalajahi M. A review on synthesis and applications of dendrimers. *J Iran Chem Soc*. 2020;18(3):503-517. doi: 10.1007/s13738-020-02053-3
 15. Alshammari RA, Aleanizy FS, Aldarwesh A, et al. Retinal delivery of the protein kinase C- β inhibitor ruboxistaurin using non-invasive nanoparticles of polyamidoamine dendrimers. *Pharmaceutics*. 2022;14(7):1444. doi: 10.3390/pharmaceutics14071444
 16. Wang J, Li B, Kompella UB, Yang H. Dendrimer and dendrimer gel-derived drug delivery systems: Breaking bottlenecks of topical administration of glaucoma medications. *MedComm Biomater Appl*. 2023;2(1):e30. doi: 10.1002/mba2.30
 17. López-Cano JJ, González-Cela-Casamayor MA, Andrés-Guerrero V, Herrero-Vanrell R, Molina-Martínez IT. Liposomes as vehicles for topical ophthalmic drug delivery and ocular surface protection. *Expert Opin Drug Deliv*. 2021;18(7):819-847. doi: 10.1080/17425247.2021.1872542
 18. Tasharrofi N, Nourozi M, Marzban A. How liposomes pave the way for ocular drug delivery after topical administration. *J Drug Deliv Sci Technol*. 2021;67:103045. doi: 10.1016/j.jddst.2021.103045
 19. Joy JM, Amruth P, Rosemol Jacob M, Dara PK, Renuka V, Anandan R. Liposome mediated encapsulation and role of chitosan on modulating liposomal stability to deliver potential bioactives-a review. *Food Hydrocoll Health*. 2023;4:100142. doi: 10.1016/j.fhfh.2023.100142
 20. Lai S, Chen J, Zhou K, et al. Liposomes for effective drug delivery to the ocular posterior chamber. *J Nanobiotechnology*. 2019;17(1):64. doi: 10.1186/s12951-019-0498-7
 21. Zych M, Wojnar W, Kaczmarczyk-Sedlak I, Folwarczna J, Kielanowska M. Effect of berberine on glycation, aldose reductase activity, and oxidative stress in the lenses of streptozotocin-induced diabetic rats *in vivo*-a preliminary study. *Int J Mol Sci*. 2020;21(12):4278. doi: 10.3390/ijms21124278
 22. Kim SK, Kang H, Ban JY, Park SI. Anti-apoptotic effect of chrysophanol isolated from *Cassia tora* seed extract on blue-light-induced A2E-loaded human retinal pigment epithelial cells. *Int J Mol Sci*. 2023;24(7):6676. doi: 10.3390/ijms24076676
 23. Sunil DK, Pradhan R, Hejmady S, et al. Emerging innovations in nano-enabled therapy against age-related macular degeneration: A paradigm shift. *Int J Pharm*. 2021;600:120499. doi: 10.1016/j.ijpharm.2021.120499
 24. Choradiya BR, Patil SB. A comprehensive review on nanoemulsion as an ophthalmic drug delivery system. *J Mol Liq*. 2021;339:116751. doi: 10.1016/j.molliq.2021.116751
 25. Fernandes AR, Souto EB, Santos TD, Silva AM, Garcia ML, Sanchez-Lopez E. Development and characterization of nanoemulsions for ophthalmic applications: Role of cationic surfactants. *Materials (Basel)*. 2021;14(24):7541. doi: 10.3390/ma14247541
 26. Pardeshi SR, Jain RS, Rajput RL, et al. Development and optimization of sustained release moxifloxacin hydrochloride loaded nanoemulsion for ophthalmic drug delivery: A 3² factorial design approach. *Micro Nanosys*. 2021;13(3):292-302. doi: 10.2174/1876402912999200826111031
 27. Youssef AAA, Thakkar R, Dudhipala N, Joshi PH, Majumdar S, Senapati S. Design of topical moxifloxacin mucoadhesive nanoemulsion for the management of ocular bacterial infections. *Pharmaceutics*. 2022;14(6):1246. doi: 10.3390/pharmaceutics14061246
 28. Mehroodish S, Mirzaeei S. Design of novel nanoemulsion formulations for topical ocular delivery of itraconazole: Development, characterization and *in vitro* bioassay. *Adv Pharm Bull*. 2021;12(1):93-101. doi: 10.34172/apb.2022.009
 29. Kassem AA, Salama A, Mohsen AM. Formulation and optimization of cationic nanoemulsions for enhanced ocular delivery of dorzolamide hydrochloride using box-behnken design: *In vitro* and *in vivo* assessments. *J Drug Deliv Sci Technol*. 2021;68:103047.
 30. Walenga RL, Babiskin AH, Zhang X, Absar M, Zhao L, Lionberger RA. Impact of vehicle physicochemical properties on modeling-based predictions of cyclosporine ophthalmic emulsion bioavailability and tear film breakup time. *J Pharm Sci*. 2018;108(1):620-629. doi: 10.1016/j.xphs.2018.10.034
 31. Altamimi MA, Imam SS, Hussain A, Alshehri S, Alnemer UA. Development and evaluations of transdermally delivered luteolin loaded cationic nanoemulsion: *In vitro* and *ex vivo* evaluations. *Pharmaceutics*. 2021;13(8):1218. doi: 10.3390/pharmaceutics13081218
 32. Jurišić Dukovski B, Juretić M, Bračko D, et al. Functional ibuprofen-loaded cationic nanoemulsion: Development and optimization for dry eye disease treatment. *Int J Pharm*. 2019;576:118979. doi: 10.1016/j.ijpharm.2019.118979
 33. Agarwal P, Rupenthal ID, Craig JP. Formulation considerations for the management of dry eye disease. *Pharmaceutics*. 2021;13(2):207. doi: 10.3390/pharmaceutics13020207
 34. Shih KC, Chan TC, Lam PY, Jhanji V, Fong PY, Tong L. Role of tear film biomarkers in the diagnosis and management of dry eye disease. *Taiwan J Ophthalmol*. 2019;9(3):150. doi: 10.4103/tjo.tjo_56_19
 35. Grobbelaar M, Louw GE, Sampson SL, Van Helden PD, Donald PR, Warren RM. Evolution of rifampicin treatment for tuberculosis. *Infect Genet Evol*. 2019;74:103937. doi: 10.1016/j.meegid.2019.103937
 36. Bazán Henostroza MA, Curo Melo KJ, Nishitani Yukuyama M, Löbenberg R, Araci Bou-Chacra N. Cationic rifampicin nanoemulsion for the treatment of ocular tuberculosis. *Colloids Surf A Physicochem Eng Asp*. 2020;597:124755. doi: 10.1016/j.colsurfa.2020.124755
 37. Dhiman N, Awasthi R, Kulkarni GT, Sharma B, Kharkwal H. Lipid nanoparticles as carriers for bioactive delivery. *Front Chem*. 2021;9:580118. doi: 10.3389/fchem.2021.580118
 38. Jacob S, Shan J, Nair AB, et al. Lipid nanoparticles as a promising drug delivery carrier for topical ocular therapy-an overview on recent advances. *Pharmaceutics*. 2022;14(3):533. doi: 10.3390/pharmaceutics14030533
 39. Mu H, Holm R. Solid lipid nanocarriers in drug delivery: Characterization and design. *Expert Opin Drug Deliv*. 2018;15(8):771-785. doi: 10.1080/17425247.2018.1504018
 40. Sastri KT, Radha GV, Pidikiti P, Vajjhala P. Solid lipid nanoparticles: Preparation techniques, their characterization, and an update on recent studies. *J Appl Pharm Sci*. 2020;10(6):126-141. doi: 10.7324/japs.2020.10617

41. Khames A, Khaleel MA, El-Badawy MF, El-Nezhawy AOH. Natamycin solid lipid nanoparticles - sustained ocular delivery system of higher corneal penetration against deep fungal keratitis: Preparation and optimization. *Int J Nanomedicine*. 2019;14:2515-2531. doi: 10.2147/ijn.s190502
42. Masoumi A, Soleimani M, Azizkhani M, et al. Clinical features, risk factors, and management of candida keratitis. *Ocul Immunol Inflamm*. 2023;32(7):1169-1174. doi: 10.1080/09273948.2023.2203752
43. Manikandan P, Abdel-Hadi A, Singh YRB, et al. Fungal keratitis: Epidemiology, rapid detection, and antifungal susceptibilities of *Fusarium* and *Aspergillus* isolates from corneal scrapings. *Biomed Res Int*. 2019;2019:6395840. doi: 10.1155/2019/6395840
44. Khan S, Sharma A, Jain V. An overview of nanostructured lipid carriers and its application in drug delivery through different routes. *Adv Pharm Bull*. 2022;13(3):446-460. doi: 10.34172/apb.2023.056
45. Lakhani P, Patil A, Wu KW, et al. Optimization, stabilization, and characterization of amphotericin B loaded nanostructured lipid carriers for ocular drug delivery. *Int J Pharm*. 2019;572:118771. doi: 10.1016/j.ijpharm.2019.118771
46. Regueiro U, López-López M, Varela-Fernández R, Sobrino T, Diez-Feijoo E, Lema I. Immunomodulatory effect of human lactoferrin on toll-like receptors 2 expression as therapeutic approach for keratoconus. *Int J Mol Sci*. 2022;23(20):12350. doi: 10.3390/ijms232012350
47. Varela-Fernández R, García-Otero X, Díaz-Tomé V, et al. Lactoferrin-loaded nanostructured lipid carriers (NLCs) as a new formulation for optimized ocular drug delivery. *Eur J Pharm Biopharm*. 2022;172:144-156. doi: 10.1016/j.ejpb.2022.02.010
48. Deka M, Ahmed AB, Chakraborty J. Development, evaluation and characteristics of ophthalmic *in situ* gel system: A review. *Int J Curr Pharm Res*. 2019;11(4):47-53. doi: 10.22159/ijcpr.2019v11i4.34949
49. Paul S, Majumdar S, Chakraborty M. Revolutionizing ocular drug delivery: Recent advancements in *in situ* gel technology. *Bull Natl Res Cent*. 2023;47(1):154. doi: 10.1186/s42269-023-01123-9
50. Huang H, Qi X, Chen Y, Wu Z. Thermo-sensitive hydrogels for delivering biotherapeutic molecules: A review. *Saudi Pharm J*. 2019;27(7):990-999. doi: 10.1016/j.jsps.2019.08.001
51. Li Y, Wang M, Wang F, Lu S, Chen X. Recent progress in studies of photocages. *Smart Mol*. 2023;1(1):e2022003. doi: 10.1002/smo.20220003
52. Pandey M, Choudhury H, Binti Abd Aziz A, et al. Potential of stimuli-responsive *in situ* gel system for sustained ocular drug delivery: Recent progress and contemporary research. *Polymers (Basel)*. 2021;13(8):1340-1340. doi: 10.3390/polym13081340
53. Vigani B, Rossi S, Sandri G, Bonferoni MC, Caramella CM, Ferrari F. Recent advances in the development of *in situ* gelling drug delivery systems for non-parenteral administration routes. *Pharmaceutics*. 2020;12(9):859. doi: 10.3390/pharmaceutics12090859
54. Majeed A, Khan NA. Ocular *in situ* gel: An overview. *J Drug Deliv Ther*. 2019;9(1):337-347. doi: 10.22270/jddt.v9i1.2231
55. Wei Y, Li C, Zhu Q, Zhang X, Guan J, Mao S. Comparison of thermosensitive *in situ* gels and drug-resin complex for ocular drug delivery: *In vitro* drug release and *in vivo* tissue distribution. *Int J Pharm*. 2020;578:119184. doi: 10.1016/j.ijpharm.2020.119184
56. Mahboobian MM, Mohammadi M, Mansouri Z. Development of thermosensitive *in situ* gel nanoemulsions for ocular delivery of acyclovir. *J Drug Deliv Sci Technol*. 2020;55:101400. doi: 10.1016/j.jddst.2019.101400
57. Wang X, Ye X, Zhang Y, Ji F. Flurbiprofen suppresses the inflammation, proliferation, invasion and migration of colorectal cancer cells via COX2. *Oncol Lett*. 2020;20(5):132. doi: 10.3892/ol.2020.11993
58. Prahladbhai Patel A, Patel JK. Mucoadhesive *in-situ* gel formulation for vaginal delivery of tenofovir disoproxil fumarate. *Indian J Pharm Educ Res*. 2020;54(4):963-970. doi: 10.5530/ijper.54.4.190
59. Yurtdaş-Kırımlioğlu G. A promising approach to design thermosensitive *in situ* gel based on solid dispersions of desloratadine with kolliphor® 188 and pluronic® F127. *J Therm Anal Calorim*. 2021;147(2):1307-1327. doi: 10.1007/s10973-020-10460-0
60. Ranch KM, Maulvi FA, Naik MJ, Koli AR, Parikh RK, Shah DO. Optimization of a novel *in situ* gel for sustained ocular drug delivery using box-behnken design: *In vitro*, *ex vivo*, *in vivo* and human studies. *Int J Pharm*. 2019;554:264-275. doi: 10.1016/j.ijpharm.2018.11.016
61. Wang L, Pan W, Pan H, et al. A novel carbon dots/thermo-sensitive *in situ* gel for a composite ocular drug delivery system: Characterization, *ex-vivo* imaging, and *in vivo* evaluation. *Int J Mol Sci*. 2021;22(18):9934. doi: 10.3390/ijms22189934
62. Wei W, Cao H, Shen D, Sun X, Jia Z, Zhang M. Antioxidant carbon dots nanozyme loaded in thermosensitive *in situ* hydrogel system for efficient dry eye disease treatment. *Int J Nanomedicine*. 2024;19:4045-4060. doi: 10.2147/ijn.s456613
63. Li S, Tang Y, Zhang X, Dou Y, Shen X. Preparation and characterization of diclofenac sodium β -cyclodextrin inclusion complex eye drops. *J Incl Phenom Macrocycl Chem*. 2019;94(1-2):85-94. doi: 10.1007/s10847-019-00910-0
64. Gorantla S, Waghule T, Rapalli VK, et al. Advanced hydrogels based drug delivery systems for ophthalmic delivery. *Recent Pat Drug Deliv Formul*. 2020;13(4):291-300. doi: 10.2174/1872211314666200108094851
65. Xu H, Liu Y, Jin L, et al. Preparation and characterization of ion-sensitive brimonidine tartrate *in situ* gel for ocular delivery. *Pharmaceutics (Basel)*. 2023;16(1):90. doi: 10.3390/ph16010090
66. Sun J, Sun X. Preparation of a novel tacrolimus ion sensitive ocular *in situ* gel and *in vivo* evaluation of curative effect of immune conjunctivitis. *Pharm Dev Technol*. 2022;27(4):399-405. doi: 10.1080/10837450.2022.2067870
67. Al-Kinani AA, Zidan G, Elsaid N, Seyfoddin A, Alani AWG, Alany RG. Ophthalmic gels: Past, present and future. *Adv Drug Deliv Rev*. 2017;126:113-126. doi: 10.1016/j.addr.2017.12.017
68. Xie M, Wang H, Gao T, et al. The protective effect of luteolin on the depression-related dry eye disorder through sirt1/NF- κ B/NLRP3 pathway. *Aging (Albany NY)*. 2023;15(1):261-275. doi: 10.18632/aging.204479
69. Omran S, Elnaggar YSR, Abdallah OY. Carrageenan tethered ion sensitive smart nanogel containing oleophytocubosomes for improved ocular luteolin delivery. *Int J Pharm*. 2023;646:123482. doi: 10.1016/j.ijpharm.2023.123482
70. Zan M, Lin L, Xu H, Zhang X. Self-Assembled Lyotropic Liquid Crystals Nanoparticles Systems for A-Arbutin Protection and Skin Delivery. [Preprint]; 2024. doi: 10.12139/ssrn.4955466
71. Ma Q, Luo R, Zhang H, et al. Design, characterization, and application of a ph-triggered *in situ* gel for ocular delivery of vinpocetine. *AAPS Pharm SciTech*. 2020;21(7):253. doi: 10.1208/s12249-020-01791-0
72. Kouchak M, Mahmoodzadeh M, Farrahi F. Designing of a pH-triggered carbopol®/hpmc *in situ* gel for ocular delivery of dorzolamide HCl: *In vitro*, *in vivo*, and *ex vivo* evaluation. *AAPS PharmSciTech*. 2019;20(5):210. doi: 10.1208/s12249-019-1431-y
73. Barse RK, Tagalpallewar AA, Kokare CR, Sharma JP, Sharma PK. Formulation and *ex vivo-in vivo* evaluation of pH-triggered brimonidine tartrate *in situ* gel for the glaucoma treatment using application of 3² factorial design. *Drug Dev Ind Pharm*. 2018;44(5):800-807. doi: 10.1080/03639045.2017.1414229
74. Bharath S, Karuppaiah A, Siram K, Hariharan S, Santhanam R. Development and evaluation of a pH triggered *in situ* ocular gel of brimonidine tartrate. *J Res Pharm*. 2020;24(3):416-424.

- doi: 10.35333/jrp.2020.164
75. Yokoyama Y, Kawasaki R, Takahashi H, et al. Effects of brimonidine and timolol on the progression of visual field defects in open-angle glaucoma: A single-center randomized trial. *J Glaucoma*. 2019;28(7):575-583. doi: 10.1097/jjg.0000000000001285
 76. Jayaram H, Kolko M, Friedman DS, Gazzard G. Glaucoma: Now and beyond. *Lancet*. 2023;402(10414):1788-1801. doi: 10.1016/s0140-6736(23)01289-8
 77. Sakr MG, El-Zahaby SA, Al-Mahallawi AM, Ghorab DM. Fabrication of betaxolol hydrochloride-loaded highly permeable ocular bilosomes (HPOBs) to combat glaucoma: *In vitro*, *ex vivo* & *in vivo* characterizations. *J Drug Deliv Sci Technol*. 2023;82:104363. doi: 10.1016/j.jddst.2023.104363
 78. Hu J, Li H, Zhao Y, et al. Critical evaluation of multifunctional betaxolol hydrochloride nanoformulations for effective sustained intraocular pressure reduction. *Int J Nanomedicine*. 2022;17:5915-5931. doi: 10.2147/ijn.s382968
 79. Allam A, Elsbahy M, El Badry M, Eleraky NE. Betaxolol-loaded niosomes integrated within pH-sensitive *in situ* forming gel for management of glaucoma. *Int J Pharm*. 2021;598:120380. doi: 10.1016/j.ijpharm.2021.120380
 80. Durak S, Esmaili Rad M, Alp Yetisgin A, et al. Niosomal drug delivery systems for ocular disease-recent advances and future prospects. *Nanomaterials (Basel)*. 2020;10(6):1191. doi: 10.3390/nano10061191
 81. Verma A, Tiwari A, Saraf S, Panda PK, Jain A, Jain SK. Emerging potential of niosomes in ocular delivery. *Expert Opin Drug Deliv*. 2020;18(1):55-71. doi: 10.1080/17425247.2020.1822322
 82. Tian Y, Zhang T, Li J, Tao Y. Advances in development of exosomes for ophthalmic therapeutics. *Adv Drug Deliv Rev*. 2023;199:114899. doi: 10.1016/j.addr.2023.114899
 83. Cao X, Xue LD, Di Y, Li T, Tian YJ, Song Y. MSC-derived exosomal lncRNA SNHG7 suppresses endothelial-mesenchymal transition and tube formation in diabetic retinopathy via MiR-34a-5p/XBP1 axis. *Life Sci*. 2021;272:119232. doi: 10.1016/j.lfs.2021.119232
 84. Kalam MA, Iqbal M, Alshememry A, Alkholief M, Alshamsan A. Fabrication and characterization of tedizolid phosphate nanocrystals for topical ocular application: Improved solubilization and *in vitro* drug release. *Pharmaceutics*. 2022;14(7):1328. doi: 10.3390/pharmaceutics14071328
 85. Lawson HD, Walton SP, Chan C. Metal-organic frameworks for drug delivery: A design perspective. *ACS Appl Mater Interfaces*. 2021;13(6):7004-7020. doi: 10.1021/acsami.1c01089
 86. Gupta C, Upreti S, Punya, Singh J, Ghosh MP, Basu T. Rapid electrochemical quantification for *in vitro* release trait of ophthalmic drug loaded within mucoadhesive metal organic framework (MOF). *ChemistrySelect*. 2021;6(12):3006-3012. doi: 10.1002/slct.202004558
 87. He Y, Ye Z, Liu X, et al. Can machine learning predict drug nanocrystals? *J Control Release*. 2020;322:274-285. doi: 10.1016/j.jconrel.2020.03.043
 88. German C, Chen Z, Przekwas A, et al. Computational model of *in vivo* corneal pharmacokinetics and pharmacodynamics of topically administered ophthalmic drug products. *Pharm Res*. 2023;40(4):961-975. doi: 10.1007/s11095-023-03480-6
 89. Delpierre C, Lefèvre T. Precision and personalized medicine: What their current definition says and silences about the model of health they promote. Implication for the development of personalized health. *Front. Sociol*. 2023;8:1112159. doi: 10.3389/fsoc.2023.1112159
 90. Mitchell MJ, Billingsley MM, Haley RM, Wechsler ME, Peppas NA, Langer R. Engineering precision nanoparticles for drug delivery. *Nat Rev Drug Discov*. 2020;20(1):101-124. doi: 10.1038/s41573-020-0090-8
 91. Tetyczka C, Brisberger K, Reiser M, et al. Itraconazole nanocrystals on hydrogel contact lenses via inkjet printing: Implications for ophthalmic drug delivery. *ACS Appl Nano Mater*. 2022;5(7):9435-9446. doi: 10.1021/acsanm.2c01715

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