

Empowering of novel anti-tumor formulations with quinone-based active natural products

Xiao Xiao^{1,2}, Baozhong Duan², Yuan Zhuang^{1,3*}, and Lei Zhang^{1,2,3*}

ABSTRACT

Cancer remains one of the leading causes of death worldwide, representing a significant threat to human health. Consequently, there is an urgent need to develop effective treatment strategies with low toxicity. Quinone-based natural products have garnered considerable attention in the field of anticancer research due to their distinctive chemical structures. These compounds play a crucial role in treating various cancers and in overcoming chemotherapy resistance through several mechanisms, including the inhibition of cell proliferation and migration, as well as the modulation of multiple signaling pathways. However, their clinical application is limited by severe side effects, which arise from certain physicochemical properties, such as poor water solubility and low biocompatibility. The advent of nanotechnology has led to the development of nanomedicine delivery systems, offering a groundbreaking approach to overcome these limitations. Nanocarriers, characterized by their excellent biocompatibility, favorable pharmacokinetics, and high drug-loading capacities, enhance the bioavailability and targeting of natural products while reducing adverse effects. Therefore, integrating quinone-based natural products with nanocarrier delivery systems has proven to be an effective anticancer strategy. This approach not only improves the absorption of drugs with poor bioavailability but also significantly reduces side effects. Various nanodelivery systems, including micelles, liposomes, inorganic nanoparticles, and biomimetic nanocarriers, are particularly effective in delivering quinone-based natural products due to their unique physical and chemical properties, thereby enhancing their solubility and stability. In addition, targeted modifications, intelligent controlled release, and combination therapy strategies have significantly improved their bioavailability and antitumor efficacy. This review systematically examines the antitumor potential of quinone-based natural products and provides a comprehensive overview of the current research and clinical application prospects of their nanodelivery systems in cancer treatment. It aims to summarize the current progress and clinical prospects of integrating these compounds with nanocarrier-based drug delivery systems in cancer treatment.

Keywords:

Drug delivery; Nanotechnology; Natural products; Quinones

*Corresponding authors:

Yuan Zhuang,
zhuangyuan@zstu.edu.cn;
Lei Zhang,
idrc@zstu.edu.cn

How to cite this article:

Xiao X, Duan B, Zhuang Y, Zhang L. Empowering of novel anti-tumor formulations with quinone-based active natural products. *Biomater Transl.* 2025

doi: [10.12336/bmt.24.00089](https://doi.org/10.12336/bmt.24.00089)



1. Introduction

Quinones are naturally occurring compounds found extensively in plants, fungi, bacteria, and animals.¹ These compounds either possess an intramolecular unsaturated cyclic diketone structure or can readily transform into such a structure.² Structurally, natural quinones are primarily classified into four categories: benzoquinone, naphthoquinone, phenanthrenequinone, and anthraquinone.³ The presence of chromophores, such as –OH

and –OCH₃, imparts vivid colors such as yellow, red, and purple to these molecules. Quinones and their derivatives are integral components of several widely used traditional Chinese medicines (TCMs), including *Rhei radix et rhizome*, *Polygoni cuspidati rhizoma et radix*, *Salviae miltiorrhizae radix et rhizoma*, and *Arnebiae radix*, among others.

Natural benzoquinones are categorized into two types: Ortho-benzoquinone and para-benzoquinone, as depicted in **Figure 1**. The

Natural quinone-based antitumor formulations

structure of ortho-benzoquinone tends to be unstable due to steric hindrance, making para-benzoquinone the more prevalent natural form. On the other hand, the majority of naturally occurring naphthoquinones are derivatives of alpha-naphthoquinone, exemplified by shikonin (SHK) and isoshikonin derivatives found primarily in the TCM *Arnebiae radix*. Meanwhile, phenanthrenequinone derivatives, such as tanshinone I, tanshinone IIA (TSIIA), dihydrotanshinone, and cryptotanshinone, are extractable from the roots of *S. miltiorrhiza* Bunge, a prominent TCM source of phenanthrenequinones.⁴ Finally, anthraquinones are classified based on their nuclear structure into monoanthracene and bianthracene types, including substances such as rhein, emodin, alizarin, aloin, and hypericin.

Extensive research, both domestic and international, has highlighted the diverse biological activities of quinones,⁵⁻¹¹ including antitumor, anti-inflammatory, antibacterial, antiviral, antifungal, and antimalarial properties, as well as neuroprotective effects against Alzheimer's disease. These biological activities position quinones as significant agents in medicinal chemistry,¹² offering promising prospects for the development of medical applications.¹³ Quinone-based natural products can function as anticancer agents through various mechanisms, such as inducing cancer cell death by inhibiting quinone oxidoreductase 1 (NQO1), leading to reactive oxygen species (ROS) generation, and inhibiting tumor growth through the suppression of signal transducer and activator of transcription 3 (STAT3).¹²

Despite the extensive documentation of their anticancer potential, the clinical application of these compounds is often hampered by challenges such as low solubility, poor absorption, rapid metabolism, and low bioavailability *in vivo*. To overcome these limitations, substantial research has been conducted on various nanomaterial drug delivery systems that are suitable for quinone-active compounds, aiming to enhance the targeting capability and bioavailability of these drugs.

Nanotechnology offers a promising alternative to traditional treatment methods by creating efficient and safe targeted delivery systems through the encapsulation of drugs within nanoparticles.^{14, 15} This review provides a comprehensive overview of current methodologies and key reference points for optimizing the formulation of quinone-based natural products using various drug delivery systems to enhance efficacy (Figure 2). It compiles and summarizes both classical and recent studies, highlighting the biological activities and potential limitations of nanomaterials. The aim is to offer valuable insights into nanodelivery systems for quinone-based natural products, with a specific focus on enhancing their anticancer efficacy.

2. Research methodology

An extensive electronic search was conducted across prominent databases, including PubMed, ScienceDirect, CNKI, Web

of Science, and Google Scholar. The search also extended to clinical trial studies listed on ClinicalTrials.gov. Strategies of the search involved identifying key terms, such as "Quinone," "benzoquinone," "naphthoquinone," "phenanthrenequinone," "anthraquinone," "structure," "bioactivity," "anti-tumor," "nanotechnology," "bioavailability," and "anti-cancer." These terms were specifically searched within the "Title/Abstract/Keywords" fields of the databases, with no restrictions on publication date. The primary aim of this comprehensive search was to identify and retrieve all relevant published studies encompassing extensive experimental designs, *in vitro* and *in vivo* analyses, clinical trials, and case-control studies. These studies primarily focused on investigating the role of nanotechnology in advancing the development, formulation, and therapeutic applications of quinone-based natural products for antitumor therapy. Special attention was given to factors such as improved bioavailability, targeted tumor delivery, enhanced cellular uptake, controlled drug release, and overall advancements in cancer treatment. In addition, information on the efficacy of preclinical models and results that have already been applied in clinical settings were also the focus. Duplicate publications were excluded from the review, and the findings regarding the antitumor activities of quinones were systematically organized within the relevant sections.

3. Antitumor activity and mechanisms of action of quinones

Quinone compounds are ubiquitously present in nature and can be synthesized by a variety of organisms.¹⁶ This widespread availability provides a rich resource base for the exploration and application of quinones. The plethora of natural products not only facilitates the identification and development of quinone compounds with potential antitumor properties but also supports their large-scale production and clinical utilization. Quinone compounds exhibit diverse antitumor mechanisms, targeting multiple cellular components and signaling pathways, thus offering broad therapeutic possibilities. Extensive research has investigated the use of quinones and their analogs in treating various cancers, including those of the lung, liver, breast, colon, cervix, prostate, and leukemia.¹⁷⁻¹⁹ At the molecular level, the anticancer efficacy of quinones is attributed to several mechanisms, such as the inhibition of cell proliferation and migration, the induction of apoptosis, and the modulation of autophagy and other cellular pathways.^{20, 21} Some quinones disrupt cell cycle progression, thereby inhibiting cell proliferation. For instance, thymoquinone (TQ) impedes the activity of cyclin-dependent kinases, arresting human breast and colon cancer cells in the G1 phase.²² In addition, emodin targets the HER-2/neu tyrosine kinase activity, consequently inhibiting the proliferation of breast cancer cells.²³

Tumor growth and metastasis depend on neovascularization, which provides nutrients and pathways for invasion. Quinone

¹Innovative Drug Research Center, College of Life Sciences and Medicine, Zhejiang Sci-Tech University, Hangzhou, Zhejiang, China; ²Chinese Medicine Resource Center, College of Pharmaceutical Science, Dali University, Dali, Yunnan, China; ³Department of Pharmaceutical Botany, School of Pharmacy, Naval Medical University, Shanghai, China

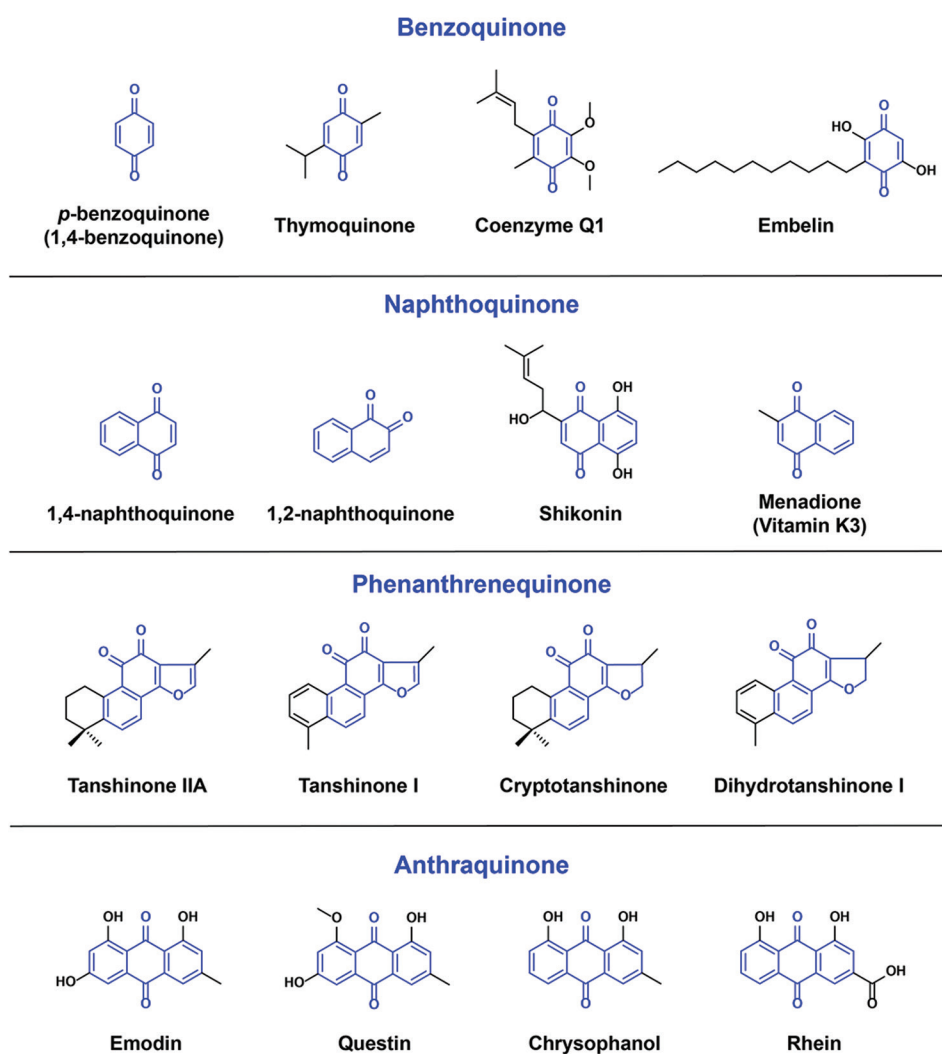


Figure 1. Representative structures of quinones from different categories, including natural products derived from quinones and synthetic quinones used as drugs

compounds counteract tumor angiogenesis by inhibiting vascular endothelial growth factor (VEGF) and its signaling pathways. For example, research by Xie *et al.*²⁴ demonstrated that TSIIA inhibits angiogenesis in the A549 human non-small cell lung cancer cell line by targeting the protein kinase domain of VEGF/VEGFR2, thereby impeding tumor growth and metastasis. Quinones also promote tumor cell apoptosis through various mechanisms.²⁵⁻²⁷ A significant pathway involves the regulation of cellular redox homeostasis and the mediation of ROS production. Lu *et al.*²⁸ discovered that SHK enhances the expression of receptor-interacting protein kinase 1 (RIPK1) and RIPK3, leading to the excessive accumulation of intracellular ROS and mitochondrial superoxide in a concentration-dependent manner. This accumulation forms a positive feedback loop with RIP1 and RIP3, triggering necroptosis in glioma cells, with ROS playing a central role in the expression of RIPK1 and RIPK3 and the assembly of necrotic bodies. Similarly, research by Liu *et al.*²⁹ suggests that ROS acts as an upstream signal for RIPK1 and RIPK3, mediating not only SHK-induced necrotic apoptosis in nasopharyngeal carcinoma cells but also regulating the expression of the RIPK1/RIPK3/MLKL complex, thereby

highlighting the pivotal role of ROS in quinone-induced necrotic apoptosis. In addition, quinones can mitigate tumor chemotherapy resistance by inhibiting the overexpression of ATP-binding cassette transporters and modulating multiple signaling pathways.^{30, 31} This inhibition enhances the efficacy of chemotherapy drugs and synergistically combats tumors in conjunction with other chemotherapeutic agents.

Quinone compounds, encompassing various types such as benzoquinone, naphthoquinone, phenanthrenequinone, and anthraquinone, feature a core structure with conjugated double bonds and active functional groups, including carbonyl groups. This distinct structural configuration confers substantial chemical reactivity upon them. In the context of structure-activity relationships, quinones such as SHK, tanshinones, and emodin are increasingly recognized as potential candidates for anticancer drug development. Despite the absence of quinone-based natural products being independently utilized as anticancer agents in current clinical practice, ongoing clinical registration studies are likely to expedite their translation into clinical applications. For instance, an early clinical trial demonstrated that an SHK derivative could inhibit lung cancer growth and was both safe and effective.^{32, 33} In addition,

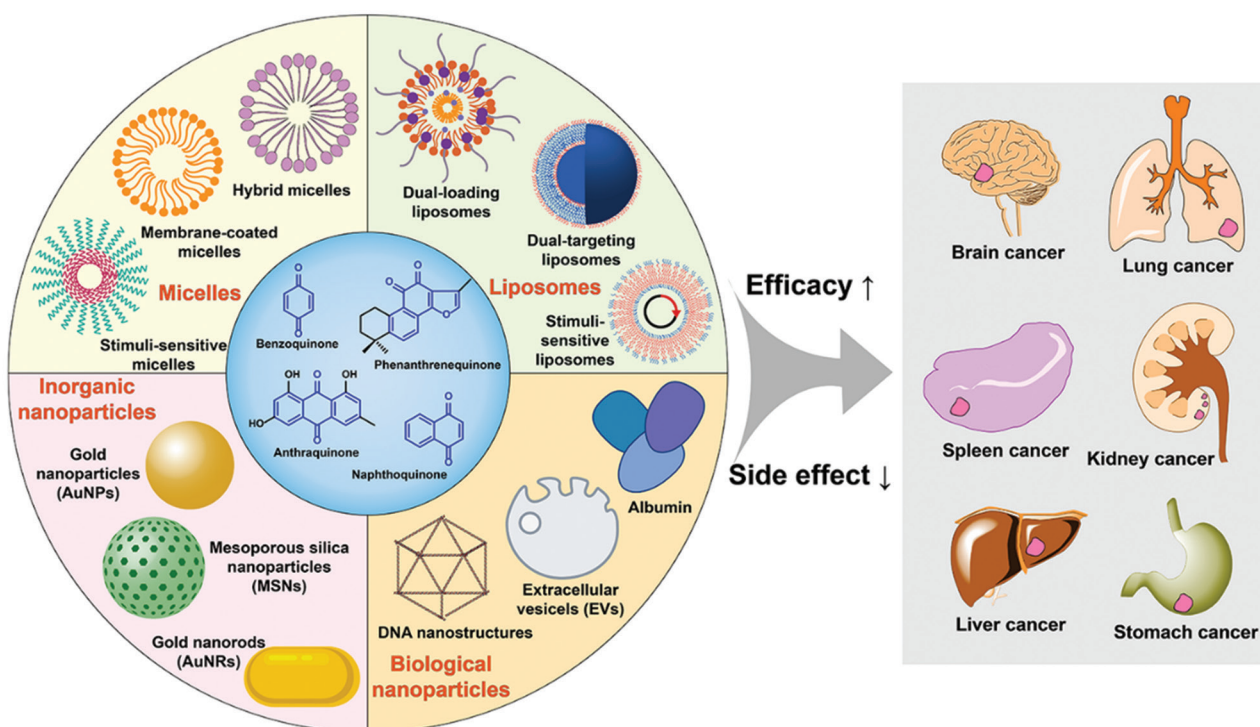


Figure 2. A series of nanomaterials that could be used in quinone formulations for precise delivery and treatment of various types of cancer

SHK has also been registered for clinical trial studies on the treatment of bladder urothelial carcinoma and breast cancer.³⁴

Benzoquinone, a class of quinones featuring a single six-membered ring,³⁵ exists primarily in two isomeric forms: 1,4-benzoquinone and 1,2-benzoquinone.³⁶ The more prevalent 1,4-benzoquinone, with the chemical formula $C_6H_4O_2$, is an oxidized derivative of 1,4-hydroquinone and is commonly referred to as para-benzoquinone (or *p*-benzoquinone).³⁵ It forms the basic unit of ubiquinone (coenzyme Q),³⁷ playing a crucial role as an electron carrier in various bioenergetic processes, including phosphorylation and electron transport within mitochondria.³⁸ Furthermore, it has demonstrated significant anticancer potential. For instance, TQ, an active constituent extracted from the black seed (*Nigella sativa* L.), has been extensively investigated as an anticancer agent.³⁹ It exerts its effects by arresting proliferation and mitosis through the stabilization of microtubules, inducing G2/M cell cycle arrest, inhibiting histone deacetylase and human telomerase reverse transcriptase activity, promoting histone acetylation and autophagy, and reducing the phosphorylation of protein kinase B (AKT) and S6 kinase beta-1. Moreover, the combined use of TQ with paclitaxel (PTX) or doxorubicin (Dox) has shown enhanced antitumor activity compared to the use of each agent individually.⁴⁰ Notably, pre-treatment with TQ has been found to sensitize 4T1 cells to PTX, resulting in increased cytotoxicity.⁴¹

Naphthoquinones, naturally occurring as 1,4-naphthoquinone (para-isomer) and 1,2-naphthoquinone (ortho-isomer), are characterized by a C6 – C4 skeleton.⁴² These compounds are categorized based on their nuclear structures into three types: “1,4-naphthoquinones,” “1,2-naphthoquinones,” and “2,6-naphthoquinones”, with 1,4-naphthoquinones being

the most prevalent. The 1,4-naphthoquinones are noted for their ability to alkylate tumor cells through the electrophilic arylation of their core structure and the potent reducibility of hydroxyl groups. As electrophilic agents, naphthoquinones interact with electronegative substances within cells, thereby manifesting pharmacological activities.⁴³ Among these, SHKs are particularly noteworthy. Extracted from the Boraginaceae family, an important group in TCM, SHKs possess a 5,8-dihydroxy-1,4-naphthoquinone structure with an isohexenyl side chain.⁴⁴ Research has consistently shown that SHKs exert robust anticancer properties across various cancer types by inhibiting cell proliferation and migration, as well as inducing apoptosis, autophagy, and necroptosis. In addition, SHKs promote ROS generation, suppress exosome release, and enhance antitumor immunity through modulation of the phosphoinositide 3-kinase (PI3K)/AKT/mammalian target of rapamycin (mTOR) and mitogen-activated protein kinase signaling pathways. They also inhibit the activity of proteins such as thioredoxin reductase 1, pyruvate kinase muscle isozyme, PIP1/3, proto-oncogene tyrosine-protein kinase Src, and focal adhesion kinase, and regulate the expression of endoplasmic reticulum protein 57, matrix metalloproteinases, activating transcription factor 2, C-Myc proto-oncogene, miR-128, and glucose-regulated protein 78 (binding immunoglobulin protein).⁴⁵ SHKs can overcome drug resistance mechanisms in tumor cells, induce programmed cell death (necroptosis), and demonstrate efficacy against both drug-sensitive and drug-resistant tumor cells. These cells often exhibit high expression of proteins such as P-glycoprotein, B-cell lymphoma 2, and B-cell lymphoma-extra large, and SHKs show significant antitumor activity both *in vitro* and *in vivo*. Yang *et al.*⁴⁶ identified that the interaction of carbonyl

carbons C1 and C4 of SHK with the tumor proteasome contributes to the suppression of proteasomal activity and the induction of cell death *in vivo*. Furthermore, menadiione (vitamin K3), another naturally occurring 1,4-naphthoquinone lacking 5,8-hydroxy groups, induces classical oxidative stress-induced necrosis but not necroptosis.⁴⁷ Xuan *et al.*⁴⁸ noted that the presence of 5,8-hydroxy groups in the 1,4-naphthoquinone nucleus is crucial for inducing necroptosis when comparing naphthoquinone derivatives of SHK with vitamin K3.

Phenanthrenequinones, another quinonoid compound, bind to DNA molecules to exert cytotoxic effects.⁴⁹ Tanshinones, a prominent class of phenanthrenequinones, are the principal compounds extracted from the dried roots and rhizomes of *S. miltiorrhiza* Bge.⁵⁰ The structure of tanshinones underlies their cytotoxic capabilities.^{51,52} Their molecular configuration, which includes furan rings and quinone structures, generates free radicals and induces DNA damage, thereby inhibiting DNA synthesis in tumor cells. This leads to alterations in the expression of numerous genes associated with tumor cell proliferation, differentiation, apoptosis, and invasion. It also affects the expression of cell surface antigens and reverses multidrug resistance.⁵³ TSIIA, the most studied lipophilic component of *S. miltiorrhiza*, significantly inhibits tumor proliferation, disrupts the cell cycle, and induces apoptosis through the PI3K/AKT/mTOR and the c-Jun N-terminal kinase signaling pathways. In addition, it promotes autophagic death and inhibits cell migration and invasion.⁵⁴

The core structure of anthraquinones consists of a rigid, planar three-ring aromatic anthracene system, featuring two keto groups at the 9- and 10-positions. The planar anthraquinone core can integrate into the DNA double helix of cancer cells, where it participates in a specific redox cycle. This cycle generates the superoxide radical anion ($O_2^{\cdot-}$) *in vivo*, leading to the interruption of DNA replication and the induction of DNA coding errors.⁵⁵ Anthraquinones disrupt the proliferation and differentiation of tumor cells, promote apoptosis, obstruct the cell cycle, modulate immune signaling, alter cell migration, and counteract multidrug resistance in the progression of malignant tumors.⁵⁶ For instance, emodin, a natural derivative of anthraquinone found in widely used TCMs such as *Rheum palmatum*, *Polygonum cuspidatum*, and *Polygonum multiflorum*,⁵⁷ has demonstrated significant therapeutic efficacy against various tumors.⁵⁸ Researchers have designed and synthesized numerous anthraquinone derivatives targeting diverse cancer types, with several drugs, including mitoxantrone,⁵⁹ Dox,⁶⁰ and epirubicin,⁶¹ already approved for clinical use.

In summary, distinct quinone compounds exhibit unique antitumor mechanisms and hold potential for drug development.³⁵ Benzoquinones, with their relatively simple structures, are readily amenable to chemical modification. By introducing various substituent groups, their antitumor activity and pharmacokinetic properties can be effectively enhanced. Despite their high chemical reactivity, which may contribute to pronounced cytotoxicity and adverse reactions, these compounds remain of interest. On the other hand, naphthoquinones primarily induce cell apoptosis or necrosis through the promotion of ROS accumulation⁶²

and ferroptosis.⁶³ Their high redox activity, however, may result in significant cytotoxicity and oxidative stress damage. Phenanthrenequinones, noted for their unique DNA embedding abilities, also exhibit anti-inflammatory properties,⁶⁴ which are beneficial for improving the tumor microenvironment. Nonetheless, their pharmacokinetic attributes require further optimization. Anthraquinones, recognized for their substantial antitumor efficacy, are among the most extensively studied quinone compounds. Despite their widespread clinical use, issues such as cardiotoxicity and drug resistance persist. Moreover, the generally poor water solubility of quinone compounds limits their stability and bioavailability *in vivo*. Thus, strategic approaches involving structural optimization, targeted modification, and nanodelivery systems can significantly enhance the antitumor efficacy of various quinone compounds, reduce their toxicity, and augment their clinical value, thereby fostering their advanced development in clinical applications.

4. Development of nanotechnology-based quinone delivery systems

Quinone-based compounds have demonstrated substantial antitumor efficacy *in vitro*, yet their clinical application is hindered by several challenges, including low stability, poor water solubility, erratic pharmacokinetics, and a lack of targeting specificity. For instance, SHK and its derivatives, despite their significant therapeutic potential, are plagued by issues related to pharmacokinetics, toxicity, and stability, such as poor bioavailability, nephrotoxicity, and susceptibility to photodegradation.^{65,66} Similarly, the natural hydrophobic compound TSIIA, exhibits a brief half-life ($t_{1/2}$) of only 44 min⁶⁷ and a low oral bioavailability of approximately 2.9 – 3.4% in rats.⁶⁸ Furthermore, the water solubility of emodin is limited to a mere 70 mg/L,⁶⁹ while plumbagin is constrained by a median elimination half-life and a mean residence time of 9.6 and 5.0 h, respectively.⁷⁰

Nanotechnology has emerged as a novel cancer treatment strategy, particularly as an efficient delivery system with significant therapeutic potential. Nanocarriers, characterized by their excellent biocompatibility, favorable pharmacokinetics, and high drug-loading capacity, enhance the bioavailability and targeting precision of natural products while diminishing side effects.⁷¹⁻⁷³ Nanomaterials typically range in size from 1 to 100 nm and exhibit unique optical, magnetic, and electrical properties, rendering them optimal for biomedical applications.⁷⁴ Over the past two decades, nanomaterials have garnered significant interest in the biomedical domain, prompting the development of various nanodelivery systems designed to amplify the anticancer efficacy of quinone-based natural products (**Figure 3**). **Table 1** provides a compilation of anticancer quinone-based drugs utilizing a diverse array of nanomaterials, including micelles, lipid-based systems, dendrimers, and inorganic and biological nanomaterials.

4.1. Micelles

Polymeric micelles consist of amphiphilic polymers that form structures with an internal hydrophobic core, which entraps

Natural quinone-based antitumor formulations

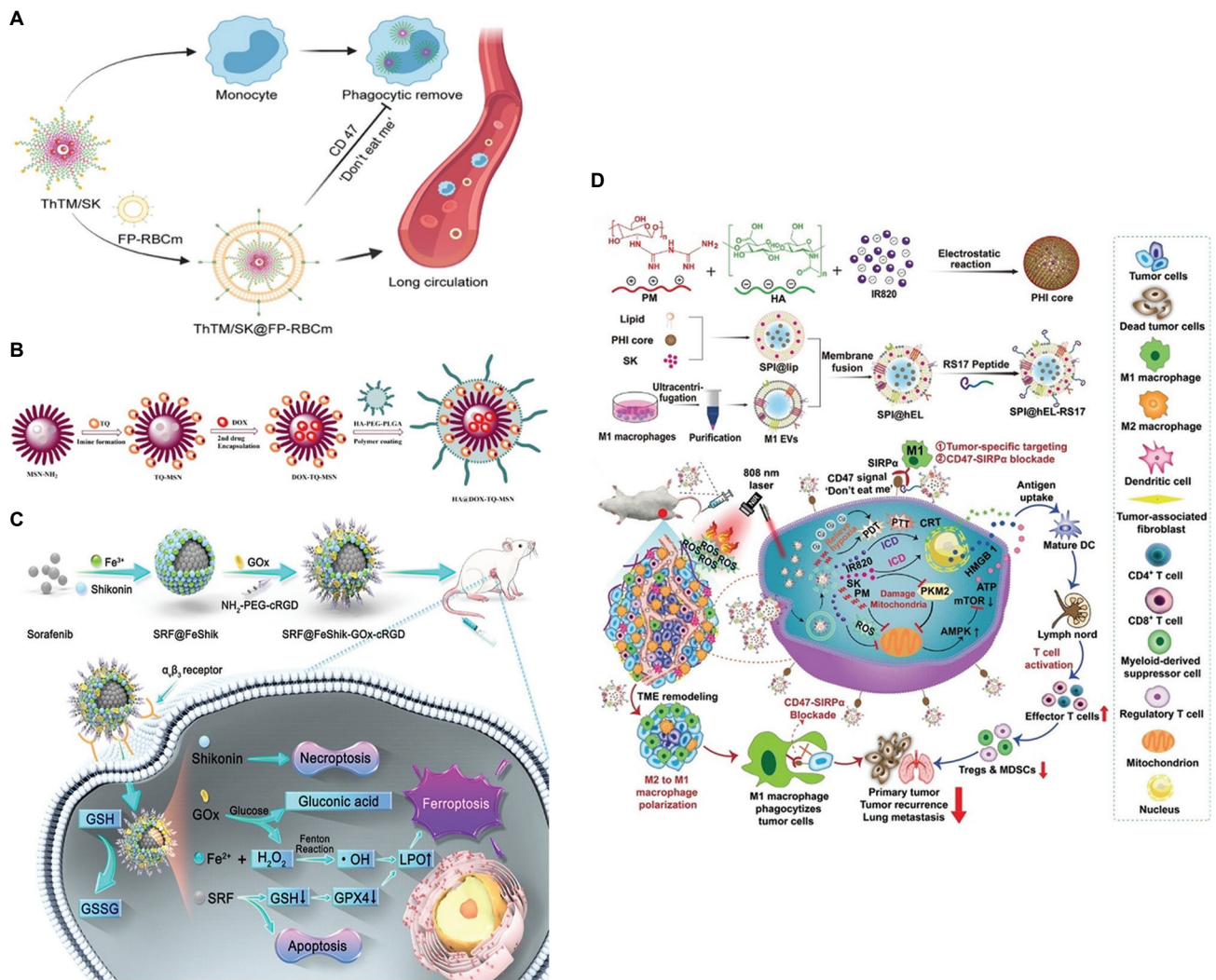


Figure 3. Schematic illustrating several nanomaterials-based intracellular delivery systems for the accurate delivery of quinones and quinone derivatives. (A) The effect of micelles *in vivo*, including two scenarios: with and without RBCm camouflage. Micelles with RBCm camouflage (ThTM/SK@FP-RBCm) can evade phagocytic cells and exhibit prolonged circulation compared to those without RBCm camouflage. Reprinted from Peng *et al.*⁷⁶ Image used with permission from Wiley-VCH GmbH, Copyright © 2022, Wiley-VCH GmbH. (B) The amine functional groups on the side-chains of MSNs react with the carbonyl groups of TQ to form TQ-MSN. Subsequently, DOX is added to generate DOX-TQ-MSN. Finally, hyaluronic acid-polyethylene glycol-poly(lactic-co-glycolic acid) (HA-PEG-PLGA) is used for encapsulation, resulting in a simple, effective, and pH-responsive drug delivery system (HA@DOX-TQ-MSN). Reprinted from Bhattacharjee *et al.*¹¹⁷ Image used with permission from American Chemical Society, Copyright © 2024, American Chemical Society. (C) Fe(III)-Shikonin (FeShik) is used as a carrier to internally load sorafenib (SRF) and externally attach GOx and amine-polyethylene glycol-cyclo(Arg-Gly-Asp-d-Phe-Lys) (NH₂-PEG-cRGD). FeShik releases Fe²⁺ and consumes GSH in tumor cells overexpressing GSH, thereby inducing tumor cell death through iron deposition. GOx promotes the production of •OH by providing an acidic environment and a high concentration of H₂O₂. Following the decomposition of SNs, the released SRF inhibits GSH biosynthesis, leading to the inactivation of GPX4 and enhancing the iron deposition effect for tumor treatment. Reprinted from Feng *et al.*¹²³ Image used with permission from American Chemical Society, Copyright © 2022, American Chemical Society. (D) PM interacts with polyanions, multivalent charged HA, and the photosensitizer IR820 via electrostatic reactions to form a kernel formed by PM, HA, and IR820 together (PHI) core. This water-soluble PHI core is loaded into the hydrophilic cavity of liposomes, while the hydrophobic drug shikonin (SK) is accommodated in its phospholipid bilayer, resulting in the formation of SPI@lip. SPI@lip then fuses with M1 macrophage extracellular vesicles (M1 EV) to form hybrid nanovesicles, SPI@hEL. The RS17 peptide is subsequently incorporated into SPI@hEL using post-insertion technology, resulting in SPI@hEL-RS17 nanoparticles (NPs). Reprinted from Tang *et al.*⁷⁷ Image used with permission from Wiley-VCH GmbH, Copyright © 2023, Wiley-VCH GmbH.

Abbreviations: AMPK: Adenosine 5'-monophosphate-activated protein kinase; CD4/8/47: Cluster of differentiation 4/8/47; CRT: Calreticulin; DC: Dendritic cell; GSSG: Glutathione disulfide; HMGB1: High mobility group protein 1; ICD: Immunogenic cell death; LPO: Lipid peroxides; MDSC: Myeloid-derived suppressor cell; mTOR: Mammalian target of rapamycin; PKM2: Pyruvate kinase M2; ROS: Reactive oxygen species; SIRPα: Signal-regulatory protein alpha; TME: Tumor microenvironment; Tregs: Regulatory T cells; RBCm: Red blood cell membrane; MSNs: Mesoporous silica nanoparticle; TQ: Thymoquinone; DOX: Doxorubicin; GOx: Glucose oxidase; SNs: Supramolecular nanomedicines; GPX4: Glutathione peroxidase 4; PM: Polymetformin; HA: Hyaluronic acid.

Table 1. Examples of anti-cancer quinone drugs based on a variety of nanomaterials

Natural product	Formulation	Materials for delivery platform	Cancer type	Characteristics	References
Shikonin	STP-NG/SHK	<i>L</i> -phenylalanine <i>N</i> -carboxyanhydride, <i>L</i> -cystine <i>N</i> -carboxyanhydride, amino-terminated mPEG, trifluoroacetic acid, and tert-butoxycarbonyl-amino-modified PEG-NH ₂	Osteosarcoma	Mean particle size: 85.90±5.50 nm, Zeta potential: -2.21 mV	75
	ThTM/SK@FP-RBCm	NH ₂ -PEG _{2k} -PCL _{6.6k} , NH ₂ -PEG _{3.4k} -Hyd-PCL _{6.6k} , mPEG _{2k} -PCL _{6.6k} , and FA-PEG-FA (PEG _{1k} , PEG _{2k} , and PEG _{3.4k})	Triple-negative breast cancer	Mean particle size: 140.00 nm	76
	SPI@hEL-RS17	Polymetformin, new indocyanine green, hyaluronic acid, M1 macrophages	4T1 breast tumor, B16F10 melanoma	Mean particle size: 131.90±4.00 nm, Zeta potential: -21.68±0.96 mV	77
Emodin	SNM EO;	Mesoporous propylamine;	Colon cancer	Mean particle size: 250.00±80.00 nm;	78
	SNM-M EO;	Mesoporous propylamine and <i>N</i> -methyl isatoic anhydride;		230.00±90.00 nm;	
	SNM-L EO	Mesoporous propylamine and lissamine rhodamine B sulfonyl chloride		260.00±80.00 nm	
	MLE	1,2-dioleoylsn-glycero-3-phosphoethanolamine- <i>n</i> - [poly (ethyleneglycol)]; ferromagnetic iron oxide nanocubes	Breast cancer	Mean particle size: 136.30 nm, Zeta potential: -18.10 mV	79
	E-SLNs	Glycerol monostearate, stearic acid, Tween 80, Poloxamer 188	Breast cancer	Mean particle size: 28.60±3.10 nm, Zeta potential: -17.00 – -24.00 mV	80
Tanshinone IIA	MSN-FA-TAN-MB	Mesoporous silica nanoparticle, microbubbles	Hepatocellular carcinoma, cervical carcinoma, lung cancer	-	81
	Gal-pH-TanIIA-NSVs	Non-ionic surfactant vesicles, galactosylated stearate	Liver cancer	Mean particle size: 53.72±0.91 nm, Zeta potential: -28.31±1.44 mV	82
	MSN-TanIIA-PEG	Polyethylenimine-polyethylene glycol, mesoporous silica	Liver cancer	Mean particle size: 117.00 nm, Zeta potential: 43.20 mV	83

Abbreviations: FA: Folic acid; mPEG: Methoxypolyethylene glycols; PCL: Polycaprolactone; PEG: Polyethylene glycol; STP-NG: Sarcoma-targeting peptide-decorated disulfide-crosslinked polypeptide nanogel.

poorly water-soluble drugs, and an external hydrophilic shell that insulates the encapsulated drugs.⁸⁴ Recent advancements in micelle-based drug delivery systems have markedly enhanced the therapeutic efficacy, specificity, and safety of quinone-based bioactive natural products in cancer treatment. These systems utilize innovative strategies, including targeted delivery, reactive drug release, and combination therapy, to optimize the anticancer potential of quinone compounds. Functionalized micelles, such as those incorporating glycyrrhetic acid (GA) and triphenylphosphine, facilitate tumor-specific and mitochondria-targeted delivery. Chen *et al.*⁸⁵ reported the development of a novel redox-responsive micellar system (GA-PEG-SS-PLGA) with GA-mediated hepatoma-targeting capabilities. This system has been shown to effectively deliver TSIIA for hepatocellular carcinoma (HCC) therapy, resulting in elevated liver TSIIA accumulation, targeted delivery, enhanced bioavailability, increased pro-apoptotic effects, and robust antitumor efficacy both *in vitro* and *in vivo*. Furthermore, Peng *et al.*⁷⁶ developed an innovative “right-side-out” red blood cell

(RBC) membrane-coated cationic micelle system (ThTM/SK@FP-RBCm) specifically for the treatment of triple-negative breast cancer (TNBC). This system enhances the mitochondria-targeted delivery of SHK facilitated by triphenylphosphine while also reducing non-specific serum binding, immune recognition, and rapid cationic nanoparticle elimination in circulation. The strategy demonstrated enhanced tumor accumulation of the drugs, effective mitochondrial biogenesis inhibition through DNA polymerase subunit gamma downregulation, and significant antitumor and antimetastatic effects, with high safety and potential applicability for other mitochondria-dependent cancers (**Figure 3A**).

Stimuli-responsive micelles, encompassing pH-, ROS-, and temperature-sensitive systems, significantly enhance drug delivery by improving solubility, stability, and controlled release. Su *et al.*⁸⁶ developed thermosensitive nanomicelles (STNs) through reversible deactivation radical polymerization to enhance the solubility, tumor targeting, and therapeutic

Natural quinone-based antitumor formulations

efficacy of SHK in breast cancer treatment. This was achieved by leveraging temperature-regulated drug release and passive targeting mechanisms, resulting in enhanced cellular uptake, cytotoxicity, and *in vivo* tumor accumulation. It was observed that STNs at 40°C facilitated rapid internalization and release of SHK into the cytoplasm of breast cancer cells, yielding a heightened cytotoxic effect compared to STNs at 37°C and free SHK (**Figure 4A**). Cheng *et al.*⁸⁷ engineered a pH-responsive and cluster of differentiation 44 (CD44)-targeting polymer micelle (Emo@CD44p-PM), self-assembled from a CD44 peptide-conjugated polyethylene glycol-block-hydroxyethyl starch-block-poly(*L*-lactic acid) (CD44p-conjugated PEG-b-HES-b-PLA). This formulation enhances the stability, cellular uptake, and antitumor efficacy of emodin by utilizing an acid-labile acetal bond. Under acidic tumor environments (pH 6.5), this bond triggers the detachment of the PEG layer and the exposure of the CD44-targeting peptide, facilitating targeted breast cancer therapy (**Figure 4B**). Furthermore, Hu *et al.*⁸⁸ developed ROS-responsive micelles (SHK@HA-PBAP) for targeted ovarian cancer therapy. These micelles, loaded with

SHK through hyaluronic acid-phenylboronic acid pinacol ester conjugation (HA-PBAP), degrade in high ROS environments to release SHK and quinone methide, simultaneously inducing ROS production, depleting intracellular glutathione (GSH), and disrupting redox homeostasis to effectively eradicate tumor cells. Liang *et al.*⁸⁹ designed and synthesized ROS/pH dual-sensitive dendrimer polymer micelles (p[*m*PEG-co-HPBE-co-EMD]@CLB) for the co-delivery of emodin and chlorambucil. These micelles were prepared with methoxypolyethylene glycols (mPEG) as the hydrophilic end and p-(hydroxymethyl) phenylboronic acid pinacol ester (HBPE) and emodin as the hydrophobic end, enabling tumor-targeted drug release through mechanisms of oxidative stress induction, GSH depletion, and sustained drug release through ROS generation and pH-triggered structural disruption at tumor sites. In addition, a GSH-responsive, lipolic acid-inulin-based delivery system was developed for TSIIA targeting colorectal cancer cells.⁹⁰ This system not only enhances drug release in tumor environments but also exhibits potent anticancer effects and supports *Bifidobacterium longum* growth, thereby merging

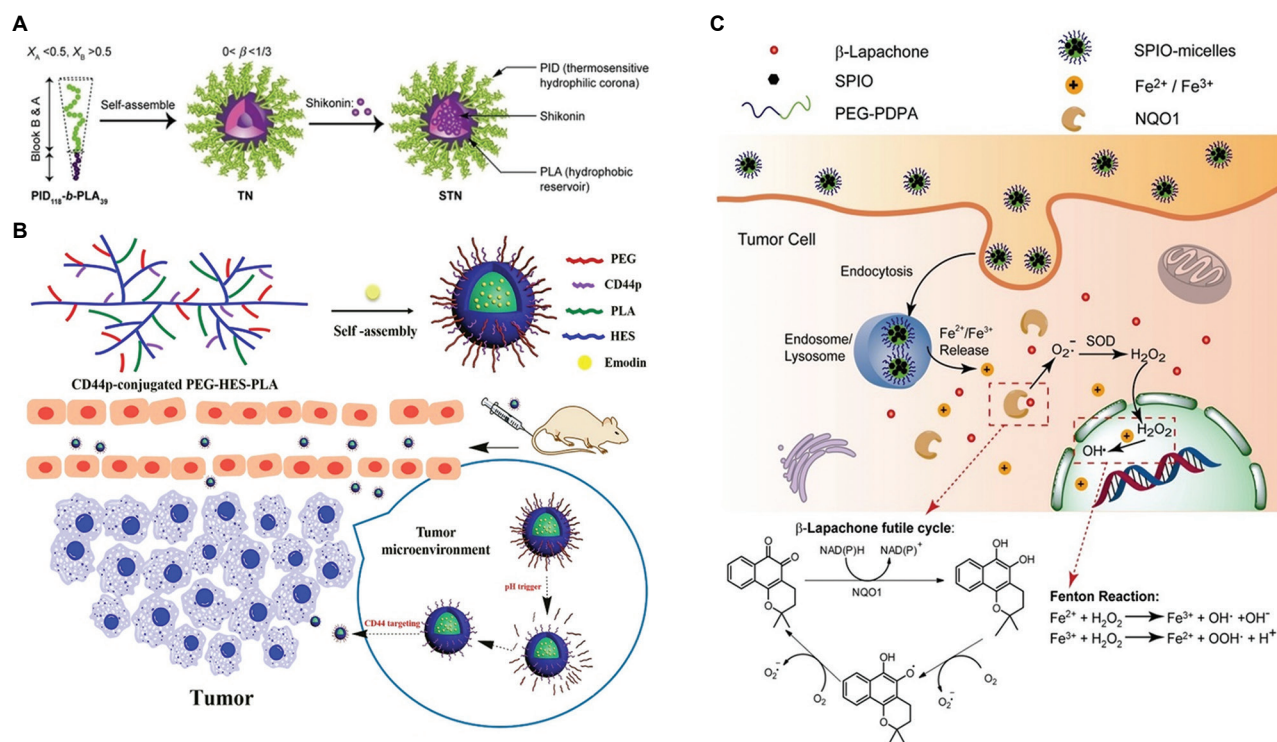


Figure 4. Micellar systems enhance the anticancer activity of quinone-based natural products. (A) The self-assembly of shikonin (SHK)-loaded thermosensitive nanomicelles (STN) in water. Reprinted from Su *et al.*⁸⁶ Image used with permission from Dove Medical Press, Copyright © 2017, Dove Medical Press. (B) Polymer micelles (Emo@CD44p-PM) are formed by the self-assembly of amphiphilic block copolymers (PEG-b-HES-b-PLA), which consist of the cluster of differentiation 44 (CD44)-targeting peptide CD44p, the hydrophobic chain polylactic acid (PLA), the hydrophilic chain polyethylene glycol (PEG), and hydroxyethyl starch (HES). When exposed to a physiological environment (pH \approx 7.4), these micelles passively accumulate in tumor tissue through the enhanced permeability and retention effect and enter the acidic tumor microenvironment (pH < 6.8), causing the separation of the PEG layer and the protein crown, increasing the accumulation of micelles and enhancing the antitumor effect. Reprinted from Cheng *et al.*⁸⁷ Image used with permission from IOP Publishing, Copyright © 2022, IOP Publishing. (C) The pH-sensitive superparamagnetic iron oxide nanoparticles (SPION) micelles act as intracellular iron donors in cancer cells, degrading and releasing iron ions in acidic organelles. Subsequently, they react with hydrogen peroxide generated by the ineffective redox cycle of β -lapachone to form highly active hydroxyl radicals, increasing reactive oxygen species (ROS) stress in cancer cells. Reprinted from Huang *et al.*⁹¹ Image used with permission from Ivyspring International Publisher, Copyright © 2013, Ivyspring International Publisher. Abbreviations: CD44p: CD44 peptide; NAD(P)H: Quinone oxidoreductase 1; PEG-PDPA: PEG-poly(2-[diisopropylamino] ethyl methacrylate); PID: PNIPAM-c-DMAAm random copolymer; SOD: Superoxide dismutase; TN: Thermosensitive nanomicelle.

cancer therapy with microbiota benefits in an innovative therapeutic approach.

In another study, Huang *et al.*⁹¹ explored the synergistic effects of pH-responsive superparamagnetic iron oxide nanoparticle micelles with β -lapachone (β -lap) to increase therapeutic efficacy in NQO1-overexpressing cancer cells. These micelles release iron ions within acidic organelles, where they react with β -lap-generated hydrogen peroxide to amplify ROS stress, inducing cancer-specific cytotoxicity. This mechanism presents a promising theranostics nanomedicine strategy (**Figure 4C**). The ferrimagnetic mPEG-b-PHEP copolymer micelle, loaded with iron oxide nanocubes and emodin, demonstrated high heating conversion efficiency and alternating magnetic field (AMF)-activated drug release. This combined approach of magnetic hyperthermia and chemotherapy achieved over 70% tumor cell death and complete tumor elimination in mice.⁹²

Advanced systems have demonstrated the potential to enhance oral bioavailability. Hou *et al.*⁹³ developed a novel self-assembled micelle system (SHK/GFB@WGA-micelles) that effectively encapsulated SHK and gefitinib (GFB) using an innovative supercritical reverse phase evaporation method. This system was designed to improve the oral absorption of SHK and GFB by facilitating mucus penetration and enterocyte uptake, thereby significantly enhancing their bioavailability and antitumor efficacy against drug-resistant lung cancer, both *in vitro* and *in vivo*.

Hybrid micelles have broadened the scope of cancer therapy by addressing significant challenges such as drug resistance and immunosuppression. Notably, the development of valid hybrid micelles (such as SHK/siIDO1-HMs) enabled the co-delivery of SHK and indoleamine 2,3-dioxygenase 1 (IDO-1) knockdown siRNA (siIDO1), showing considerable potential for colon cancer treatment.⁹⁴ These micelles induce immunogenic cell death (ICD) and modulate IDO-1-mediated immunosuppression, suggesting a potent therapeutic approach. In addition, the pH/ROS-responsive micellar nanosystem (PPDC@ β -Lap), loaded with β -lap and camptothecin, demonstrates enhanced tumor cell uptake through charge reversal, self-amplifying drug release through ROS generation, effective multidrug resistance circumvention, and potent synergistic antitumor efficacy with low systemic toxicity *in vivo*.⁹⁵ Furthermore, a compound formulation comprising epidermal growth factor (EGF)-modified PTX micelles and EGF-modified emodin micelles has been developed for ovarian cancer.⁹⁶ This formulation demonstrates specific targeting of SKOV3 ovarian cancer cells, effectively inhibiting proliferation, invasion, and metastasis. The system exhibits significant *in vivo* pharmacodynamic results, highlighting its potential as a promising strategy for ovarian cancer treatment.

Moreover, clinically viable platforms, such as β -lap-loaded micelles, address challenges like hemolytic anemia and short circulation times, achieving effective targeting of NQO1-overexpressing tumors with reduced toxicity. Blanco *et al.*⁹⁷ established a clinically viable β -lap nanomedicine platform using polymeric micelles. This system demonstrates enhanced

tumor targeting, improved drug stability, and superior therapeutic efficacy against NQO1-overexpressing non-small cell lung cancer. The platform exhibits increased tumor accumulation, prolonged circulation, reduced hemolytic toxicity, and potent antitumor effects in both subcutaneous and orthotopic lung cancer models.

In summary, micelle-based drug delivery systems represent a promising strategy in cancer therapeutics, offering enhanced tumor targeting, controlled release, and synergistic therapeutic mechanisms. These versatile platforms significantly enhance the clinical potential of quinone-based natural products by enhancing the pharmacokinetics of hydrophobic drugs while reducing systemic toxicity. However, the effectiveness of these systems depends on critical factors, including the biological characteristics of the tumor, the specificity of the targeted ligand, and the stability of the formulation. Future research should focus on optimizing micelle-based delivery systems, expanding their clinical applications, minimizing potential side effects, and addressing challenges related to tumor heterogeneity and non-specific organ accumulation.

4.2. Lipid-based nanoparticles

Liposomes closed spherical vesicles comprising lipid bilayers, possess several appealing properties, such as superior biocompatibility, excellent biodegradability, low clearance, and minimal toxicity.⁹⁸ Recent efforts have led to the development of various delivery systems based on liposomes or lipid nanoparticles, aimed at augmenting the anticancer efficacy of quinones. For instance, Meng *et al.*⁹⁹ engineered hyaluronic acid-coated SHK liposomes (HA-SHK-Lip) specifically for the treatment of TNBC. These liposomes demonstrated enhanced drug encapsulation, targeted uptake through the CD44 receptor, increased ROS production, reduced GSH level, and significantly improved antitumor effects compared to free SHK, as observed in both *in vitro* and *in vivo* studies. Besides, Luo *et al.*¹⁰⁰ described the development of a dual-targeting liposome (CS/LyP-1-PC Lip), co-loaded with PTX and cryptotanshinone, also for TNBC chemoimmunotherapy. This formulation enhances tumor targeting through dual-receptor-mediated endocytosis, induces ICD to stimulate immune responses, and counteracts the immunosuppressive tumor microenvironment by inhibiting STAT3 activation, thereby significantly suppressing tumor growth and reducing lung metastases in TNBC models (**Figure 5A**).

In addition to improving the precision of targeting, liposome-based nanodelivery systems have alleviated the limitations associated with quinone-based natural products as pharmaceutical agents and have made advancements in the field of cancer immunotherapy. Wang *et al.*¹⁰¹ formulated a novel liposomal emodin by incorporating *D*- α -tocopheryl polyethyleneglycol 1000 succinate, achieving high encapsulation efficiency, enhanced stability, extended circulation time, improved biodistribution to the lung and kidney, and increased cytotoxicity against leukemia cells in comparison to mPEG2000-DSPE liposomal emodin. On the other hand, Chen *et al.*¹⁰² developed aloe-emodin (AE)-loaded solid lipid nanoparticles (SLNs), which notably enhanced the anticancer

tumor suppression, elevated cellular uptake, and reduced toxicity compared to pristine TQ in an orthotopic xenograft pancreatic tumor model. Similarly, Kontogiannopoulos *et al.*¹⁰⁶ successfully developed and characterized novel SHK-loaded liposomes using 1,2-dipalmitoylphosphatidylcholine (DPPC) and egg phosphatidylcholine lipids, enhancing the therapeutic index, aqueous solubility, and stability of SHK while mitigating adverse effects. They demonstrated favorable physicochemical properties, controlled-release profiles, and moderate cytotoxicity against human cancer cell lines, with DPPC liposomes showing superior drug-loading capacity and release characteristics. In their study, Eskandani and Nazemiyeh¹⁰⁷ successfully developed and characterized SHK-Act-loaded SLNs using a hot homogenization method, producing stable, spherical nanoparticles (70 – 120 nm) with high entrapment efficiency, sustained release, and enhanced fluorescent imaging properties. These nanoparticles exhibited superior *in vitro* cytotoxicity, DNA damage, and prolonged antitumor effects compared to intact SHK-Act, highlighting their potential for improved therapeutic efficacy and biodistribution studies in drug delivery research. Li *et al.*¹⁰⁸ engineered a versatile nanoparticle (SHK/siR-NPs) for co-delivering SHK and programmed death-ligand 1 (PD-L1)-knockdown siRNA, which demonstrated prolonged blood circulation and increased tumor aggregation, thereby enhancing cancer immunotherapy (Figure 5B). Wu *et al.*¹⁰⁹ constructed a laser-triggered thermosensitive metabolic nanomodulator (denoted as ISM liposome) co-encapsulating IR825-NH₂, SHK, and metformin. This approach synergized phototherapy, chemotherapy, and immunotherapy, achieving robust tumor elimination after a single injection and irradiation by amplifying oxidative stress, alleviating hypoxia, inducing ICD, and sensitizing immunosuppressive tumors to immune therapy, with minimized systemic toxicity and no observed adverse effects *in vivo* in mice.

In addition to single-drug loading, dual or multi-drug loading systems have been developed to address multidrug resistance and facilitate the concurrent administration of multiple therapies, thereby enhancing the efficacy of quinone-based treatments for cancer. For instance, a nanostructured lipid carrier that co-delivers β -lap and Dox has been demonstrated to effectively circumvent multidrug resistance in breast cancer therapy.¹¹⁰ This system increases the retention of Dox and exhibits superior anticancer activity both *in vitro* and *in vivo* compared to single-drug delivery systems. Similarly, Xie *et al.*¹¹¹ designed iron-palladium nanozyme (FePd NZ)- and SHK-encapsulated functional lipid nanoparticles (FPS-LNPs) that significantly improve cancer therapy by inducing necroptosis through ROS overexpression, overcoming resistance to apoptosis, inhibiting tumor growth, and stimulating the cytotoxic T cell immune response (Figure 5C). Meanwhile, Sakpakdeejaroen *et al.*¹¹² have reported that transferrin-conjugated lipid-polymer hybrid nanoparticles loaded with PLB effectively target tumors, achieving significant regression and elimination of B16-F10 melanoma in mice. The findings demonstrate their potential as a promising anticancer nanomedicine. Furthermore, N'Diaye *et al.*¹¹³ developed biodegradable lipid nanoparticles that co-encapsulate β -lap in the polymeric core

and the photosensitizer m-THPC in the lipid shell for combined chemo- and photodynamic therapy (PDT) of retinoblastoma. This combination shows additive cytotoxic effects and enhanced efficacy at lower doses compared to single therapies. In the research conducted by Yang *et al.*,¹¹⁴ a dual-responsive lipid nanoparticle co-delivering nitric oxide and β -lap was crafted using zinc-coordinated lipids (DSNO[Zn]) alongside helper lipids (DOPE and DSPE-PEG2000). This formulation demonstrated synergistic antitumor effects by inducing oxidative stress, disrupting redox homeostasis, damaging nuclear DNA and mitochondria, and activating caspase-3, underscoring its potential as a multifunctional tumor therapy platform. A magnetic liposomal emodin nanocomposite was developed to enhance the anticancer efficacy and targeting of emodin, achieving improved tumor cell elimination, magnetic guidance for targeted drug accumulation, and significant *in vivo* tumor suppression, highlighting its potential as a magnetically guided theranostic nanoagent.⁷⁹

In summary, liposomes represent a highly efficient delivery system for quinone-based natural products, offering advantages such as greater stability, prolonged circulation time, controlled release, and enhanced anticancer efficacy. These liposome-based delivery systems provide effective approaches to overcome challenges associated with traditional anticancer strategies, ranging from improving drug delivery to enhancing therapeutic effectiveness. These benefits contribute to the enhanced anticancer activity of quinone-based natural products by improving their pharmacokinetics and enabling targeted delivery. However, the application of these systems is currently limited by factors such as system instability, the complexity of preparation processes, and the inefficiency in targeting. Future studies are required that involve the integration of novel materials, multimodal targeting strategies, and optimization of production processes to overcome these limitations and facilitate clinical translation.

4.3. Inorganic nanoparticles

The development of inorganic nanodelivery systems for quinone-based natural products in cancer treatment has progressed through various innovative approaches. These delivery systems, which utilize inorganic nanomaterials, substantially enhance drug targeting, improve treatment efficacy, and optimize therapeutic outcomes through a range of novel methods. Concurrently, these systems aim to reduce side effects and offer a safer, more effective strategy for managing cancer. One notable study by Saeed *et al.*¹¹⁵ reported on a hybrid drug, THQG, created by chemically linking the hydrophilic compound gallic acid with the hydrophobic compound TQ. This compound was subsequently loaded onto polyethyleneimine functionalized antimonene quantum dots. The resulting drug carrier system exhibited several advantageous features, including pH-sensitive drug release, low cytotoxicity, enhanced capacity to withstand oxidative stress, capabilities for radiolabeling to analyze organ deposition, and increased efficacy compared to the individual drugs. These characteristics suggest that this nanodrug could serve as a multimodal theranostic agent in the treatment of breast and oral cancer cell lines. Similarly, gold nanoparticles conjugated with anti-EGFR antibodies and loaded with the

Natural quinone-based antitumor formulations

radiosensitizer β -lap have shown enhanced tumor targeting, pH-sensitive drug release, and improved radiotherapeutic efficacy both *in vitro* and *in vivo*.¹¹⁶ This approach presents a promising strategy for selective cancer therapy with minimal off-target effects. In a related study, a hyaluronic acid-coated mesoporous silica nanoparticle system (HA@TQ-DOX-MSN) co-delivering TQ and Dox specifically targets Dox-resistant breast cancer cells.¹¹⁷ This system reduces multidrug resistance 1 expression, enhances miR-298 levels, and overcomes drug resistance while supporting sustained pH-sensitive drug release and specific targeting of cancer cells (**Figure 3B**).

Inorganic nanocarrier drug delivery systems can significantly enhance the efficacy of quinones in cancer treatment through functional modifications or integration with various therapeutic strategies. Zhang *et al.*¹¹⁸ employed molecular docking to select TSIIA from a group of phytoestrogens as a target ligand for nanodelivery systems, incorporating modifications. Their research indicates that TSIIA-modified mesoporous silica nanoparticles offer enhanced targeted delivery to estrogen receptors, effective antitumor and antimetastatic effects, high drug-loading capacity, and reduced toxicity, presenting a promising phytoestrogen-based nanopatform for breast cancer therapy. Similarly, Zhu *et al.*⁸³ discovered that polyethyleneimine-polyethylene glycol-coated mesoporous silica nanoparticles enhance HCC treatment by delivering TSIIA in a sustained manner combined with efficient glypican-3-shRNA gene therapy. This approach improves antitumor efficacy, reduces glypican-3 expression, and provides a potential synergistic therapeutic platform. Meanwhile, Lv *et al.*⁸¹ developed a multifunctional drug delivery vehicle by combining folate-modified mesoporous silica nanoparticles with microbubbles to load TSIIA. This system integrates multiple benefits, such as high drug-loading capacity, enhanced cell membrane translocation, low cytotoxicity, ultrasound-triggered release, and tumor-imaging capabilities. It demonstrates improved cellular uptake through folate receptor-mediated endocytosis *in vitro* and significant tumor suppression *in vivo* with external ultrasound irradiation. This innovation offers a novel strategy for tumor theranostic applications. Furthermore, Ren *et al.*¹¹⁹ created the multifunctional theranostic nanopatform Fe₃O₄-PEG-Cy7-EMO, combining emodin and magnetic nanoparticles for both fluorescence and magnetic resonance imaging (MRI) and targeted therapy. The system demonstrates effective pancreatic cancer diagnosis, tumor suppression, and minimal side effects through passive targeting and synergistic therapeutic properties. Liang *et al.*¹²⁰ developed MUC1-aptamer-targeted nanoparticles (MUC1@ACS) co-delivering SHK and chitosan silver nanoparticles to induce necroptotic ICD, enhance immune cell infiltration, and suppress the growth and metastasis of TNBC through tumor-specific targeting and immune activation. In addition, Zhao *et al.*¹²¹ designed a GSH-sensitive Fe-based coordination polymer-based nanosystem functionalized with calcium peroxide (CaO₂) and β -lap as donors of O₂ and H₂O₂ for dual cascade-amplified tumor chemodynamic therapy (CDT). This system can internalize in cancer cells, release cargoes in response to cytoplasmic GSH, self-produce H₂O₂ through cyclic reactions, activate ferroptosis

and Ca²⁺ overload pathways, exacerbate tumor damage, enhance CDT efficiency, relieve hypoxic restriction, and display effective MRI capabilities. Thus, the system provides a novel tumor-initiated nano-bomb platform for dual-enhanced tumor CDT efficiency.

Supramolecular nanomedicine based on Fe-SHK has been shown to enhance therapeutic efficacy. Feng *et al.*¹²² demonstrated that Fe(III)-SHK supramolecular nanomedicine, when modified with NH₂-PEG-cRGD, improves antitumor therapy by inducing ferroptosis and necroptosis, enhancing SHK's bioavailability, reducing its toxicity, and overcoming apoptosis-induced chemoresistance. In addition, a triple ferroptosis amplification strategy incorporating Fe(III)-SHK metal-polyphenol networks, sorafenib, and glucose oxidase markedly increases oxidative stress and enhances tumor therapy through GSH-responsive effects, thereby improving antitumor efficacy through mechanisms including ferroptosis, apoptosis, and necroptosis.¹²³ It offers superior therapeutic effects for 4T1 tumors (**Figure 3C**). Likewise, a ¹⁷⁷Lu-MFeCu@Tan nanotheranostic platform was developed, combining the chemotherapy agent Tan with the radionuclide ¹⁷⁷Lu within a dual-metal nanozyme carrier MFeCu.¹²⁴ This platform enables single-photon emission computed tomography and employs a quadruple-synergistic tumor therapy approach that encompasses internal radioisotope therapy, catalysis therapy, chemotherapy, and MFeCu-mediated ferroptosis and cuproptosis therapy. Both *in vitro* and *in vivo* studies of this platform have shown significant efficacy in tumor elimination and recurrence reduction. The platform's specific response to the tumor microenvironment and high retention effect further enhance its safety, providing an effective and secure quadruple synergistic antitumor therapy, particularly beneficial for TNBC. Meanwhile, Song *et al.*⁹² developed ferrimagnetic micelles containing iron oxide nanocubes and emodin, which achieve superior magnetic hyperthermia performance, enhanced MRI contrast, and AMF-activated drug release. This system enables tumor-targeted chemotherapy and complete cancer cell elimination at low dosages, with potential applications for other hydrophobic natural products in cancer therapy. Moreover, Shi *et al.*¹²⁵ designed cytosine-guanine dinucleotide (CpG) oligodeoxynucleotide-loaded aluminum hydroxyphosphate nanoparticles coated with Fe-SHK metal-phenolic networks (Alum-CpG@Fe-SHK NPs). These personalized *in situ* nanovaccines exhibit potent antitumor effects, including the elimination of primary tumors, inhibition of distant tumors, and prevention of metastasis and recurrence. They present a viable strategy to advance personalized antitumor immunotherapy by utilizing autologous tumor cell lysates *in situ* without complex *ex vivo* processes. These diverse delivery systems offer multiple strategies, including drug combination, targeted delivery, gene therapy integration, enhanced therapeutic modalities, immunotherapy, and theranostic applications for improved cancer treatment.

Inorganic nanoparticles provide unique advantages for the antitumor application of natural products, including high stability, multifunctional synergistic therapy, and precise

controlled-release capabilities. Their multifunctionality opens avenues for therapeutic and diagnostic applications, yet challenges concerning their biological safety, preparation cost, and targeting efficiency remain. Future research and optimization of these nanoparticles are essential to maximize their effectiveness and safety in cancer treatment.

4.4. Biological nanoparticles

Ideally, a drug delivery system should remain stable within the bloodstream while effectively releasing drugs at the target site. However, exogenous nanoparticles are readily identified and eliminated by the immune system. To circumvent this, leveraging endogenous components such as proteins and cell membranes for camouflage – to evade immune detection and enhance tumor-specific uptake – presents a highly promising approach.¹²⁶ The biomimetic nanodelivery system employing quinone-based natural products in cancer therapy primarily aims to address the multifaceted challenges of cancer treatment. It integrates various therapeutic modalities, including chemotherapy, phototherapy, immunotherapy, and treatment for iron deficiency, into highly targeted and efficient formulations. Tang *et al.*⁷⁷ designed bioinspired hybrid nanovesicles (SPI@hEL-RS17) by merging extracellular vesicles from M1 macrophages with liposomes and decorating them with the RS17 peptide to target tumors. This design blocks the CD47-SIRP α checkpoint and reprograms tumor-associated macrophages into tumor-inhibiting M1-like phenotypes, thereby enhancing phagocytosis and sustaining immune responses. The co-encapsulation of SHK, the photosensitizer IR820, and the immunomodulator polymetformin provides synergistic effects in chemotherapy, phototherapy, and immunotherapy that suppress tumor growth, inhibit metastasis, and prevent recurrence in highly invasive tumor models (Figure 3D).

Similarly, to address the challenges posed by the complex preparation and adverse tumor microenvironment effects on previously reported PDT and ferroptosis-combined nanocomposites, Wu *et al.*¹²⁷ discovered that AE can induce ferroptosis by inhibiting GSH S-transferase pi 1. They prepared a biomimetic nanoplatfrom, AE@RBC/Fe nanocrystals, integrating AE as both a photosensitizer and ferroptosis activator with ferritin and RBC membranes. This enhances antitumor effects through synergistic PDT and ferroptosis, with ferritin providing oxygen and Fe³⁺, while the biomimetic modification aids in circulation and targeting. However, AE's blue light absorption limits its application in deep tumors, which suggests a future focus on both efficacy and application limitations. Zhang *et al.*¹²⁸ developed ERFe₃O₄ nanoparticles, a red-blood-cell-membrane-coated, emodin-loaded iron-based nanoplatfrom. This platform synergistically reprograms tumor-associated macrophages, reverses the tumor immune suppressive microenvironment, and enhances antitumor immunity and apoptosis by modulating lactate production and promoting M1 macrophage polarization.

In a distinct approach, Liu *et al.*¹²⁹ developed a biomimetic nanoreactor, SOD-Fe⁰@Lapa-ZRF, in which SOD-Fe⁰ and Lapa cargo were encapsulated within zeolitic imidazolate

framework 8 (ZIF-8) through self-assembly. This design facilitates tumor-targeted multi-enzyme cascade therapy, generating hydroxyl radicals (\bullet OH) that selectively eradicate cancer cells while simultaneously bolstering the antioxidant capacity of normal cells and minimizing oxidative stress-related systemic toxicity. In the study by Wang *et al.*,¹³⁰ a mannosylated lactoferrin nanoparticulate system (Man-LF NPs) was crafted for the dual-targeted biomimetic delivery of SHK and JQ1. This system effectively remodels the tumor immune microenvironment by inducing immune cell death, repolarizing tumor-associated macrophages, inhibiting glucose metabolism, and obstructing the PD-L1 checkpoint, thereby augmenting antitumor immunity and representing a promising polypharmacological approach to cancer immunotherapy. Similarly, Cui *et al.*¹³¹ developed a biomimetic nanodrug delivery system (CpG-EXO/TGM) based on endogenous exosomes for targeted brain delivery. This system co-delivers TSIIA and glycyrrhizic acid nanomicelles along with immune adjuvants, demonstrating enhanced therapeutic efficacy against glioblastoma by improving blood-brain barrier penetration, stimulating anti-glioblastoma immune responses, and preventing postoperative recurrence, thus offering a viable strategy for chemo-immunotherapy and personalized treatment. Besides, Han *et al.*¹³² designed a biomimetic nanodrug formulation (Comb-NP) that co-encapsulates PLB and dihydrotanshinone I to induce ICD in HCC cells and to reverse the immunosuppressive tumor microenvironment. This formulation enhances tumor targeting and prolongs survival in HCC mice, presenting a promising approach for chemo-immunotherapy with the potential for broader application in solid tumors.

These studies underscore the multifunctionality and potential of biomimetic nanodelivery systems to enhance the efficacy of quinone-based cancer immunotherapy by targeting the tumor microenvironment, improving drug delivery, and inducing multimodal therapeutic responses. They significantly enhance the biocompatibility of quinone-based natural products, increase cellular uptake, and improve therapeutic efficacy by mimicking biological characteristics. However, their clinical applications remain constrained by challenges related to preparation complexity, stability, and potential immunogenicity. Further optimization of biomimetic designs and in-depth evaluation of long-term toxicological properties are essential through synthetic biology techniques to advance their translational application in tumor therapy.

4.5. Others

Beyond the primary categories previously discussed, numerous scholars have further augmented the anticancer properties of quinone-based natural products through various carrier systems. For instance, Soni *et al.*¹³³ synthesized poly (DL-lactide-co-glycolide) (PLGA) nanoparticles that encapsulated TQ and PTX using a single emulsion solvent evaporation technique. These nanoparticles not only enhanced the anticancer activity but also reduced the toxic effects of PTX by decreasing the required effective dose. Likewise, Li *et al.*¹³⁴ engineered lactoferrin-modified poly(ethylene glycol)-PLGA nanoparticles for the delivery of SHK, addressing its poor aqueous solubility

and non-selective biodistribution. Their system demonstrated high drug-loading capacity, sustained release, enhanced brain targeting, prolonged circulation, and superior antiglioma efficacy compared to free SHK, underscoring its potential as a targeted delivery mechanism for glioblastoma. Peng *et al.*¹³⁵ utilized polyamidoamine (PAMAM) dendrimers and PEGylated derivatives to create acetylshikonin nanoparticles, which effectively inhibited tumors despite a reduction in antitumor activity owing to strong intermolecular forces; polyethylene glycol (PEG); PEG-PAMAM (PPG) emerged as a more stable and less cytotoxic carrier. In addition, a novel galactose-modified, pH-sensitive niosome delivery system specifically targeted HCC cells, enhanced the release and bioavailability of TSIIA, and demonstrated superior antitumor efficacy and liver specificity both *in vitro* and *in vivo*.⁸² Jiang *et al.*¹³⁶ developed emodin-loaded stearic acid-grafted chitosan oligosaccharide micelles (CSO-SA/EMO), which showed enhanced drug uptake, sustained release, and significant antitumor effects *in vitro* and *in vivo* against gastric cancer cells, presenting a promising delivery system for cancer therapy.

Furthermore, quinone-based natural products not only serve as active agents but also contribute to the structural framework of drug carriers for other therapeutics. For example, an amphiphilic carboxymethyl chitosan-rhein conjugate was designed to form micelles for the oral delivery of PTX, enhancing its bioavailability, ensuring sustained release, and demonstrating significant antitumor efficacy with low toxicity.¹³⁷ This approach suggests a potential for the oral delivery of water-insoluble drugs. Moreover, a novel PEG 3500-embelin (PEG3.5k-EB2) micelle system was developed to improve the solubility and bioavailability of embelin, facilitating its capacity to efficiently transport hydrophobic drugs like PTX and exhibiting potent antitumor activity and synergistic effects at lower doses.¹³⁸ This dual functionality highlights the significant potential of quinone-based natural products in drug therapy and opens new avenues for innovation and optimization in drug delivery systems.

5. Limitations of the study

Before concluding, it is crucial to recognize the limitations inherent in the review's design. First, the selection of specific keywords for the literature search may have excluded relevant studies that used different terminologies or presented their findings differently. Second, the focus primarily on common nanodelivery systems may have overlooked emerging or less-developed nanocarriers with unique benefits. Third, the studies included in this review varied in experimental design, as well as in the types of quinone-based natural products and nanodelivery systems investigated. Such diversity makes it difficult to directly compare results across studies and draw definitive and generalizable conclusions. Finally, although significant progress has been made in enhancing the antitumor efficacy of quinone-based natural products through nanodelivery systems, most research remains in the preclinical phase. The transition from preclinical findings to clinical applications is often complex and unpredictable, as factors such as pharmacokinetic and toxicity profiles may differ significantly

between human subjects and animal models. Therefore, further validation through rigorous clinical trials is essential to establish the efficacy and safety of these nanodelivery systems.

6. Conclusions and future perspectives

Quinone-based natural products hold considerable potential for anticancer applications but also encounter numerous challenges. Active compounds such as SHK and its derivatives, TSIIA, emodin, and PLB demonstrate promising antitumor activity *in vitro*. However, their clinical application faces challenges such as low bioavailability, poor solubility, short *in vivo* release time, and potential side effects. Considerable efforts have been made to overcome these hurdles. Nanocarriers have gained significant attention for their ability to enhance the solubility of TCMS, provide excellent stability, improve absorption, and offer better sustained-release and targeting capabilities. This review has outlined the main types of nanocarriers used in this area, including micelles, liposomes, inorganic nanocarriers, and biomimetic nanocarriers. These nanocarriers demonstrate substantial promise in enhancing the efficacy of quinone-based natural products and addressing the challenges associated with their clinical application.

Although numerous reports have explored this topic, the development of nanomedicine for delivering quinone-based natural products remains in its early stages. The limited research on quinone nanoparticles in clinical trials has slowed their clinical application, which is the ultimate goal of drug research. However, translating nanomedicine into clinical practice is a complex and lengthy process, involving significant challenges related to safety, efficacy, and production costs. Nanocarriers can introduce increased toxicity, heightened immunogenicity, higher manufacturing costs, and unforeseen risks. Consequently, the development of quinone-based nanoformulations requires conclusive evidence to establish their clinical efficacy, stability, and cost-effectiveness. To advance this field, it is crucial to conduct thorough pharmacokinetic and pharmacodynamic studies on nanocarriers containing quinone-based natural products. A deeper understanding of the interactions between nanocarriers and quinone-based natural products, along with optimization of drug release modes and timing, is critical for achieving more precise treatments. Additionally, studying the metabolism and distribution of nanodelivery systems *in vivo* is essential to minimizing side effects on normal tissues. By addressing key questions related to drug delivery, pharmacokinetics, safety, and efficacy, we aim to facilitate their clinical translation.

Although clinical research remains limited, promising results from *in vitro* studies and animal models justify further investigation into quinone-based nanoparticles as a novel approach to cancer treatment. With continuous advancements in nanomaterials and their clinical applications, new formulations of quinone-based natural products are expected to drive breakthroughs and improve therapeutic outcomes in the future.

Acknowledgement

None.

Financial support

This work was supported by the National Natural Science Foundation of China (grant no. 82225047, 82170274), the National Key Research and Development Program of China (No. 2022YFC3501703), the Natural Science Foundation of Zhejiang Province (LZ24H280003), and Basic Medical Research Project of Naval Medical University (2023QN022).

Conflicts of interest statement

The authors declare no conflicts of interest.

Author contributions

Conceptualization: YZ and LZ; *Writing – original draft:* XX; *Writing – editing & review:* All authors. All authors critically revised and edited the review and approved the final version of the manuscript.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data

Not applicable.

Open access statement

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work noncommercially, as long as appropriate credit is given and the new creations are licensed under the identical terms. Important, ancillary information, which is relevant to the parent article but cannot (and does not) appear in the print (or online) version of the journal. It may comprise additional tables, data sets, figures, movie files, audio clips, 3D structures, and other related nonessential multimedia files.

References

- Lemos TL, Monte FJ, Santos AK, et al. Quinones from plants of northeastern Brazil: Structural diversity, chemical transformations, NMR data and biological activities. *Nat Prod Res.* 2007;21(6):529-550. doi: 10.1080/14786410601130604
- Patel OPS, Beteck RM, Legoabe LJ. Antimalarial application of quinones: A recent update. *Eur J Med Chem.* 2021;210(2):113084. doi: 10.1016/j.ejmech.2020.113084
- Gomes DCN, Wellisson DSMJ, Martins DCJ. Quinones: Biosynthesis, characterization of (13) C spectroscopical data and pharmacological activities. *Chem Biodivers.* 2023;20(12):e202301365. doi: 10.1002/cbdv.202301365
- Fang WT, Deng AP, Ren ZL, et al. Research progress on quality evaluation of *Salviae Miltiorrhizae Radix et Rhizoma* (Danshen). *Zhongguo Zhong Yao Za Zhi.* 2018;43(6):1077-1085. doi: 10.19540/j.cnki.cjcm.20180109.004
- Khan MA, Tania M, Fu S, Fu J. Thymoquinone, as an anticancer molecule: From basic research to clinical investigation. *Oncotarget.* 2017;8(31):51907-51919. doi: 10.18632/oncotarget.17206
- Huang X, Jin L, Deng H, et al. Research and development of natural product tanshinone I: Pharmacology, total synthesis, and structure modifications. *Front Pharmacol.* 2022;13:920411. doi: 10.3389/fphar.2022.920411
- Siddamurthi S, Gutti G, Jana S, Kumar A, Singh SK. Anthraquinone: A promising scaffold for the discovery and development of therapeutic agents in cancer therapy. *Future Med Chem.* 2020;12(11):1037-1069. doi: 10.4155/fmc-2019-0198
- Xie Y, Shen X, Xu F, Liang X. Research progress of nano-delivery systems for the active ingredients from traditional Chinese medicine. *Phytochem Anal.* 2024;35(4):1-14. doi: 10.1002/pca.3381
- Tripathi SK, Panda M, Biswal BK. Emerging role of plumbagin: Cytotoxic potential and pharmaceutical relevance towards cancer therapy. *Food Chem Toxicol.* 2019;125(3):566-582. doi: 10.1016/j.fct.2019.01.018
- Yadav S, Sharma A, Nayik GA, et al. Review of shikonin and derivatives: Isolation, chemistry, biosynthesis, pharmacology and toxicology. *Front Pharmacol.* 2022;13:905755. doi: 10.3389/fphar.2022.905755
- Zhang P, Liu W, Wang Y. The mechanisms of tanshinone in the treatment of tumors. *Front Pharmacol.* 2023;14:1282203. doi: 10.3389/fphar.2023.1282203
- Zhang L, Zhang G, Xu S, Song Y. Recent advances of quinones as a privileged structure in drug discovery. *Eur J Med Chem.* 2021;223(15):113632. doi: 10.1016/j.ejmech.2021.113632
- Ying HZ, Yu CH, Chen HK, et al. Quinonoids: Therapeutic potential for lung cancer treatment. *Biomed Res Int.* 2020;2020:2460565. doi: 10.1155/2020/2460565
- Shi J, Kantoff PW, Wooster R, Farokhzad OC. Cancer nanomedicine: Progress, challenges and opportunities. *Nat Rev Cancer.* 2017;17(1):20-37. doi: 10.1038/nrc.2016.108
- Jahangir MA, Jain P, Verma R, et al. Transdermal nutraceuticals delivery system for CNS disease. *Cns Neurol Disord Drug Targets.* 2022;21(10):977-993. doi: 10.2174/1871527321666220112154051
- Laohavisit A, Wakatake T, Ishihama N, et al. Quinone perception in plants via leucine-rich-repeat receptor-like kinases. *Nature.* 2020;587(7832):92-97. doi: 10.1038/s41586-020-2655-4
- Luo N, Zhang K, Li X, Hu Y, Guo L. Tanshinone IIA destabilizes SLC7A11 by regulating PIAS4-mediated SUMOylation of SLC7A11 through KDM1A, and promotes ferroptosis in breast cancer. *J Adv Res.* 2024;69(15):313-327. doi: 10.1016/j.jare.2024.04.009
- Boulos JC, Rahama M, Hegazy MF, Efferth T. Shikonin derivatives for cancer prevention and therapy. *Cancer Lett.* 2019;459(20):248-267. doi: 10.1016/j.canlet.2019.04.033
- Shrimali D, Shanmugam MK, Kumar AP, et al. Targeted abrogation of diverse signal transduction cascades by emodin for the treatment of inflammatory disorders and cancer. *Cancer Lett.* 2013;341(2):139-149. doi: 10.1016/j.canlet.2013.08.023
- Zhang S, Gao Q, Li W, et al. Shikonin inhibits cancer cell cycling by targeting Cdc25s. *BMC Cancer.* 2019;19:20. doi: 10.1186/s12885-018-5220-x
- Yang L, Li J, Xu L, et al. Rhein shows potent efficacy against non-small-cell lung cancer through inhibiting the STAT3 pathway. *Cancer Manag Res.* 2019;11:1167-1176. doi: 10.2147/CMAR.S171517
- Alhmied F, Alammam A, Al Sultan B, Alshehri M, Pottoo FH. Molecular mechanisms of thymoquinone as anticancer agent. *Comb Chem High T Scr.* 2021;24(10):1644-1653. doi: 10.2174/1386207323999201027225305
- Ueno N, Kiyokawa N, Hung M. Growth suppression of low HER-2/neu-expressing breast cancer cell line MDA-MB-435 by tyrosine kinase inhibitor emodin. *Oncol Rep.* 1996;3(3):509-511. doi: 10.3892/or.3.3.509
- Xie J, Liu J, Liu H, et al. The antitumor effect of tanshinone IIA on anti-proliferation and decreasing VEGF/VEGFR2 expression on the human non-small cell lung cancer A549 cell line. *Acta Pharm Sin B.* 2015;5(6):554-563. doi: 10.1016/j.apsb.2015.07.008
- Ni M, Zhou J, Zhu Z, et al. Shikonin and cisplatin synergistically overcome cisplatin resistance of ovarian cancer by inducing ferroptosis via upregulation of HMOX1 to promote Fe²⁺ accumulation. *Phytomedicine.* 2023;112(5):154701. doi: 10.1016/j.phymed.2023.154701
- Liu D, Zhu K, Guo T, et al. Chrysophanol: A promising natural compound in cancer therapy-Mechanistic insights and future perspectives. *Pharmacol Res.* 2024;210(12):107502.

- doi: 10.1016/j.phrs.2024.107502
27. Hou Y, Zhong B, Zhao L, *et al.* A small molecule cryptotanshinone induces non-enzymatic NQO1-dependent necrosis in cancer cells through the JNK1/2/Iron/PARP/calcium pathway. *Acta Pharm Sin B.* 2025;15(2):991-1006.
doi: 10.1016/j.apsb.2024.12.005
 28. Lu B, Gong X, Wang Z, *et al.* Shikonin induces glioma cell necroptosis *in vitro* by ROS overproduction and promoting RIP1/RIP3 necrosome formation. *Acta Pharmacol Sin.* 2017;38(11):1543-1553.
doi: 10.1038/aps.2017.112
 29. Liu T, Sun X, Cao Z. Shikonin-induced necroptosis in nasopharyngeal carcinoma cells via ROS overproduction and upregulation of RIPK1/RIPK3/MLKL expression. *Oncotargets Ther.* 2019;12:2605-2614.
doi: 10.2147/OTT.S200740
 30. Wang G, Li Y, Guo Z, He Q, Liu Z, Deng B. Tanshinone I stimulates pyroptosis of cisplatin-resistant gastric cancer cells by activating the NF- κ B/Caspase-3(8)/GSDME signaling pathway. *DNA Cell Biol.* 2024;43(4):185-196.
doi: 10.1089/dna.2023.0293
 31. Wang F, Jin S, Mayca PF, *et al.* Chemical screen identifies shikonin as a broad DNA damage response inhibitor that enhances chemotherapy through inhibiting ATM and ATR. *Acta Pharm Sin B.* 2022;12(3):1339-1350.
doi: 10.1016/j.apsb.2021.08.025
 32. Guo XP, Zhang XY, Zhang SD. Clinical trial on the effects of shikonin mixture on later stage lung cancer. *Zhong Xi Yi Jie He Za Zhi.* 1991;11(10):598-9, 580.
 33. Wang Z, Yin J, Li M, *et al.* Combination of shikonin with paclitaxel overcomes multidrug resistance in human ovarian carcinoma cells in a P-gp-independent manner through enhanced ROS generation. *Chin Med.* 2019;14:7.
doi: 10.1186/s13020-019-0231-3
 34. Sun Q, Gong T, Liu M, *et al.* Shikonin, a naphthalene ingredient: Therapeutic actions, pharmacokinetics, toxicology, clinical trials and pharmaceutical researches. *Phytomedicine.* 2022;94(1):153805.
doi: 10.1016/j.phymed.2021.153805
 35. Silakari P, Priyanka, Piplani P. p-Benzoquinone as a privileged scaffold of pharmacological significance: A review. *Mini Rev Med Chem.* 2020;20(16):1586-1609.
doi: 10.2174/1389557520666200429101451
 36. Singh N, Mahmood U, Kaul VK, Gupta AP, Jirovetz L. A new alkylated benzoquinone from rhizomes of *Iris kumaonensis*. *Nat Prod Res.* 2006;20(1):75-78.
doi: 10.1080/14786410500045721
 37. Krylova NG, Kulahava TA, Cheschevik VT, Dremza IK, Semenkova GN, Zavodnik IB. Redox regulation of mitochondrial functional activity by quinones. *Physiol Int.* 2016;103(4):439-458.
doi: 10.1556/2060.103.2016.4.4
 38. Batra M, Kriplani P, Batra C, Ojha KG. An efficient synthesis and biological activity of substituted p-benzoquinones. *Bioorg Med Chem.* 2006;14(24):8519-8526.
doi: 10.1016/j.bmc.2006.08.036
 39. Aumeeruddy MZ, Mahomoodally MF. Combating breast cancer using combination therapy with 3 phytochemicals: Piperine, sulforaphane, and thymoquinone. *Cancer Am Cancer Soc.* 2019;125(10):1600-1611.
doi: 10.1002/cncr.32022
 40. Woo CC, Hsu A, Kumar AP, Sethi G, Tan KH. Thymoquinone inhibits tumor growth and induces apoptosis in a breast cancer xenograft mouse model: The role of p38 MAPK and ROS. *PLoS One.* 2013;8(10):e75356.
doi: 10.1371/journal.pone.0075356
 41. Sakalar C, Izgi K, Iskender B, *et al.* The combination of thymoquinone and paclitaxel shows anti-tumor activity through the interplay with apoptosis network in triple-negative breast cancer. *Tumour Biol.* 2016;37(4):4467-4477.
doi: 10.1007/s13277-015-4307-0
 42. Hook I, Mills C, Sheridan H. *Bioactive Naphthoquinones from Higher Plants Studies in Natural Products Chemistry.* Netherlands: Elsevier Science; 2014. p. 119-160.
 43. Wellington KW. Understanding cancer and the anticancer activities of naphthoquinones - a review. *Rsc Adv.* 2015;5(26):20309-20338.
doi: 10.1039/c4ra13547d
 44. Brockmann, H. The constitution of Alkannin, Shikonin and Alkannan. *Justus Liebigs Ann Chem.* 1936;521(1):1-47.
doi:10.1002/jlac.19365210102
 45. Guo C, He J, Song X, *et al.* Pharmacological properties and derivatives of shikonin-a review in recent years. *Pharmacol Res.* 2019;149(11):104463.
doi: 10.1016/j.phrs.2019.104463
 46. Yang H, Zhou P, Huang H, *et al.* Shikonin exerts antitumor activity via proteasome inhibition and cell death induction *in vitro* and *in vivo*. *Int J Cancer.* 2009;124(10):2450-2459.
doi: 10.1002/ijc.24195
 47. Degtrev A, Huang Z, Boyce M, *et al.* Chemical inhibitor of nonapoptotic cell death with therapeutic potential for ischemic brain injury. *Nat Chem Biol.* 2005;1(2):112-119.
doi: 10.1038/nchembio711
 48. Xuan Y, Hu X. Naturally-occurring shikonin analogues - a class of necroptotic inducers that circumvent cancer drug resistance. *Cancer Lett.* 2009;274(2):233-242.
doi: 10.1016/j.canlet.2008.09.029
 49. Wu WL, Chang WL, Chen CF. Cytotoxic activities of tanshinones against human carcinoma cell lines. *Am J Chin Med.* 1991;19(3-4):207-216.
doi: 10.1142/S0192415X91000284
 50. Jiang Z, Gao W, Huang L. Tanshinones, critical pharmacological components in *Salvia miltiorrhiza*. *Front Pharmacol.* 2019;10:202.
doi: 10.3389/fphar.2019.00202
 51. Zhang Z, Zhang J, Jin L, Song T, Wu G, Gao J. Tanshinone IIA interacts with DNA by minor groove-binding. *Biol Pharm Bull.* 2008;31(12):2342-2345.
doi: 10.1248/bpb.31.2342
 52. Szymczyk P, Majewska M, Nowak J. Proteins and DNA sequences interacting with tanshinones and tanshinone derivatives. *Int J Mol Sci.* 2025;26(2):848.
doi: 10.3390/ijms26020848
 53. Li Z, Zou J, Cao D, Ma X. Pharmacological basis of tanshinone and new insights into tanshinone as a multitarget natural product for multifaceted diseases. *Biomed Pharmacother.* 2020;130(10):110599.
doi: 10.1016/j.biopha.2020.110599
 54. Zhang W, Liu C, Li J, *et al.* Tanshinone IIA: New perspective on the anti-tumor mechanism of a traditional natural medicine. *Am J Chin Med.* 2022;50(1):209-239.
doi: 10.1142/S0192415X22500070
 55. Tian W, Wang C, Li D, Hou H. Novel anthraquinone compounds as anticancer agents and their potential mechanism. *Future Med Chem.* 2020;12(7):627-644.
doi: 10.4155/fmc-2019-0322
 56. Adnan M, Rasul A, Hussain G, *et al.* Physcion and physcion 8-O-beta-D-glucopyranoside: Natural anthraquinones with potential anticancer activities. *Curr Drug Targets.* 2021;22(5):488-504.
doi: 10.2174/1389450121999201013154542
 57. Dong X, Fu J, Yin X, *et al.* Emodin: A review of its pharmacology, toxicity and pharmacokinetics. *Phytother Res.* 2016;30(8):1207-1218.
doi: 10.1002/ptr.5631
 58. Huang Q, Lu G, Shen HM, Chung MC, Ong CN. Anti-cancer properties of anthraquinones from rhubarb. *Med Res Rev.* 2007;27(5):609-630.
doi: 10.1002/med.20094
 59. Halpern AB, Othus M, Huebner EM, *et al.* Mitoxantrone, etoposide and cytarabine following epigenetic priming with decitabine in adults with relapsed/refractory acute myeloid leukemia or other high-grade myeloid neoplasms: A phase 1/2 study. *Leukemia.* 2017;31(12):2560-2567.
doi: 10.1038/leu.2017.165
 60. Xu J, Liu D, Niu H, *et al.* Resveratrol reverses Doxorubicin resistance by inhibiting epithelial-mesenchymal transition (EMT) through modulating PTEN/Akt signaling pathway in gastric cancer. *J Exp Clin Cancer Res.* 2017;36(1):19.
doi: 10.1186/s13046-016-0487-8
 61. Liu L, Mu LM, Yan Y, *et al.* The use of functional epirubicin liposomes to induce programmed death in refractory breast cancer. *Int J Nanomedicine.* 2017;12:4163-4176.
doi: 10.2147/IJN.S133194
 62. Cao S, Li H, Ye X, *et al.* Shikonin induces the apoptosis and pyroptosis

- of EGFR-T790M-mutant drug-resistant non-small cell lung cancer cells via the degradation of cyclooxygenase-2. *Eur J Med Res.* 2024;29(1):611. doi: 10.1186/s40001-024-02187-7
63. Lu C, Zhang Z, Fan Y, Wang X, Qian J, Bian Z. Shikonin induces ferroptosis in osteosarcomas through the mitochondrial ROS-regulated HIF-1 α /HO-1 axis. *Phytomedicine.* 2024;135(14):156139. doi: 10.1016/j.phymed.2024.156139
 64. Ding B, Lin C, Liu Q, et al. Tanshinone IIA attenuates neuroinflammation via inhibiting RAGE/NF- κ B signaling pathway *in vivo* and *in vitro*. *J Neuroinflamm.* 2020;17(1):1-17. doi: 10.1186/s12974-020-01981-4
 65. Kaur K, Singh A, Sharma H, Punj S, Bedi N. Formulation strategies and therapeutic applications of shikonin and related derivatives. *Recent Adv Drug Deliv Formul.* 2022;16(1):55-67. doi: 10.2174/2667387816666220302112201
 66. Zhang F, Pan T, Wu X, Gao X, Li Z, Ren X. Non-cytotoxic doses of shikonin inhibit lipopolysaccharide-induced TNF- α expression via activation of the AMP-activated protein kinase signaling pathway. *Exp Ther Med.* 2020;20(5):45. doi: 10.3892/etm.2020.9173
 67. Jia Y, Yao D, Bi H, et al. *Salvia miltiorrhiza* Bunge (Danshen) based nano-delivery systems for anticancer therapeutics. *Phytomedicine.* 2024;128(7):155521. doi: 10.1016/j.phymed.2024.155521
 68. Yu X, Lin S, Zhou Z, et al. Role of P-glycoprotein in the intestinal absorption of tanshinone IIA, a major active ingredient in the root of *Salvia miltiorrhiza* Bunge. *Curr Drug Metab.* 2007;8(4):325-340. doi: 10.2174/138920007780655450
 69. Qiu C, Zhang JZ, Wu B, et al. Advanced application of nanotechnology in active constituents of traditional Chinese medicines. *J Nanobiotechnol.* 2023;21(1):456. doi: 10.1186/s12951-023-02165-x
 70. Sumsakul W, Plengsuriyakarn T, Na-Bangchang K. Pharmacokinetics, toxicity, and cytochrome P450 modulatory activity of plumbagin. *BMC Pharmacol Toxicol.* 2016;17(1):50. doi: 10.1186/s40360-016-0094-5
 71. Chaurasia S, Patel RR, Chaubey P, Kumar N, Khan G, Mishra B. Lipopolysaccharide based oral nanocarriers for the improvement of bioavailability and anticancer efficacy of curcumin. *Carbohydr Polym.* 2015;130(16):9-17. doi: 10.1016/j.carbpol.2015.04.062
 72. Lepeltier E, Rijo P, Rizzolio F, et al. Nanomedicine to target multidrug resistant tumors. *Drug Resist Updat.* 2020;52(5):100704. doi: 10.1016/j.drug.2020.100704
 73. Su C, Liu Y, Li R. Absorption, distribution, metabolism and excretion of the biomaterials used in Nanocarrier drug delivery systems. *Adv Drug Deliv Rev.* 2019;143(6):97-114. doi: 10.1016/j.addr.2019.06.008
 74. Gavvas S, Quazi S, Karpinski TM. Nanoparticles for cancer therapy: Current progress and challenges. *Nanoscale Res Lett.* 2021;16(1):173. doi: 10.1186/s11671-021-03628-6
 75. Li S, Zhang T, Xu W, et al. Sarcoma-targeting peptide-decorated polypeptide nanogel intracellularly delivers shikonin for upregulated osteosarcoma necroptosis and diminished pulmonary metastasis. *Theranostics.* 2018;8(5):1361-1375. doi: 10.7150/thno.18299
 76. Peng J, Hu X, Fan S, et al. Inhibition of mitochondrial biosynthesis using a "right-side-out" membrane-camouflaged micelle to facilitate the therapeutic effects of shikonin on triple-negative breast cancer. *Adv Healthc Mater.* 2022;11(18):e2200742. doi: 10.1002/adhm.202200742
 77. Tang L, Yin Y, Cao Y, et al. Extracellular vesicles-derived hybrid nanoplatforams for amplified CD47 blockade-based cancer immunotherapy. *Adv Mater.* 2023;35(35):2303835. doi: 10.1002/adma.202303835
 78. Jänicke P, Lennicke C, Meister A, Seliger B, Wessjohann LA, Kaluderović GN. Fluorescent spherical mesoporous silica nanoparticles loaded with emodin: Synthesis, cellular uptake and anticancer activity. *Mater Sci Eng C.* 2021;119(2):111619. doi: 10.1016/j.msec.2020.111619
 79. Song Y, Sheng Z, Xu Y, et al. Magnetic liposomal emodin composite with enhanced killing efficiency against breast cancer. *Biomater Sci.* 2019;7(3):867-875. doi: 10.1039/c8bm01530a
 80. Wang S, Chen T, Chen R, Hu Y, Chen M, Wang Y. Emodin loaded solid lipid nanoparticles: Preparation, characterization and antitumor activity studies. *Int J Pharmaceut.* 2012;430(1-2):238-246. doi: 10.1016/j.ijpharm.2012.03.027
 81. Lv Y, Cao Y, Li P, et al. Ultrasound-triggered destruction of folate-functionalized mesoporous silica nanoparticle-loaded microbubble for targeted tumor therapy. *Adv Healthc Mater.* 2017;6(18):1700354. doi: 10.1002/adhm.201700354
 82. Hu X, Zhang J, Deng L, Hu H, Hu J, Zheng G. Galactose-modified ph-sensitive niosomes for controlled release and hepatocellular carcinoma target delivery of tanshinone IIA. *Aaps Pharmscitech.* 2021;22(3):96. doi: 10.1208/s12249-021-01973-4
 83. Zhu Y, Yue M, Guo T, et al. PEI-PEG-coated mesoporous silica nanoparticles enhance the antitumor activity of tanshinone IIA and serve as a gene transfer vector. *Evid Based Complement Alternat Med.* 2021;2021:6756763. doi: 10.1155/2021/6756763
 84. Cagel M, Tesan FC, Bernabeu E, et al. Polymeric mixed micelles as nanomedicines: Achievements and perspectives. *Eur J Pharm Biopharm.* 2017;113(4):211-228. doi: 10.1016/j.ejpb.2016.12.019
 85. Chen F, Zhang J, He Y, Fang X, Wang Y, Chen M. Glycyrhretinic acid-decorated and reduction-sensitive micelles to enhance the bioavailability and anti-hepatocellular carcinoma efficacy of tanshinone IIA. *Biomater Sci.* 2016;4(1):167-182. doi: 10.1039/c5bm00224a
 86. Su Y, Huang N, Chen D, et al. Successful *in vivo* hyperthermal therapy toward breast cancer by Chinese medicine shikonin-loaded thermosensitive micelle. *Int J Nanomedicine.* 2017;12:4019-4035. doi: 10.2147/IJN.S132639
 87. Cheng K, Zhou J, Zhao Y, et al. pH-responsive and CD44-targeting polymer micelles based on CD44p-conjugated amphiphilic block copolymer PEG-b-HES-b-PLA for delivery of emodin to breast cancer cells. *Nanotechnology.* 2022;33(27):275604. doi: 10.1088/1361-6528/ac5f9a
 88. Hu K, Li X, Tan Z, Shi Y. Simple ROS-responsive micelles loaded Shikonin for efficient ovarian cancer targeting therapy by disrupting intracellular redox homeostasis. *Eur J Pharm Biopharm.* 2024;204(11):114525. doi: 10.1016/j.ejpb.2024.114525
 89. Liang W, Fan Y, Liu Y, et al. ROS/pH dual-sensitive emodin-chlorambucil co-loaded micelles enhance anti-tumor effect through combining oxidative damage and chemotherapy. *Int J Pharm.* 2023;647(18):123537. doi: 10.1016/j.ijpharm.2023.123537
 90. Wang D, Sun F, Lu C, et al. Inulin based glutathione-responsive delivery system for colon cancer treatment. *Int J Biol Macromol.* 2018;111(6):1264-1272. doi: 10.1016/j.ijbiomac.2018.01.071
 91. Huang G, Chen H, Dong Y, et al. Superparamagnetic iron oxide nanoparticles: Amplifying ROS stress to improve anticancer drug efficacy. *Theranostics.* 2013;3(2):116-126. doi: 10.7150/thno.5411
 92. Song Y, Li D, Lu Y, et al. Ferrimagnetic mPEG-b-PHEP copolymer micelles loaded with iron oxide nanocubes and emodin for enhanced magnetic hyperthermia-chemotherapy. *Natl Sci Rev.* 2020;7(4):723-736. doi: 10.1093/nsr/nwz201
 93. Hou X, Ai X, Liu Z, et al. Wheat germ agglutinin modified mixed micelles overcome the dual barrier of mucus/enterocytes for effective oral absorption of shikonin and gefitinib. *Drug Deliv Transl Res.* 2024;15(1):325-342. doi: 10.1007/s13346-024-01602-0
 94. Li J, Zhao M, Xu Y, Hu X, Dai Y, Wang D. Hybrid micelles codelivering shikonin and IDO-1 siRNA enhance immunotherapy by remodeling immunosuppressive tumor microenvironment. *Int J Pharm.* 2021;597(6):120310. doi: 10.1016/j.ijpharm.2021.120310

95. Dai L, Li X, Duan X, *et al.* A pH/ROS cascade-responsive charge-reversal nanosystem with self-amplified drug release for synergistic oxidation-chemotherapy. *Adv Sci.* 2019;6(4):1801807. doi: 10.1002/advs.201801807
96. Tang L, Liu XX, Yang XD, Tan S, Zou ZW. A compound formulation of EGF-modified paclitaxel micelles and EGF-modified emodin micelles enhance the therapeutic effect of ovarian cancer. *J Liposome Res.* 2023;33(1):89-101. doi: 10.1080/08982104.2022.2086568
97. Blanco E, Bey EA, Khemtong C, *et al.* Beta-lapachone micellar nanotherapeutics for non-small cell lung cancer therapy. *Cancer Res.* 2010;70(10):3896-3904. doi: 10.1158/0008-5472.CAN-09-3995
98. Malam Y, Loizidou M, Seifalian AM. Liposomes and nanoparticles: Nanosized vehicles for drug delivery in cancer. *Trends Pharmacol Sci.* 2009;30(11):592-599. doi: 10.1016/j.tips.2009.08.004
99. Meng L, Ren J, Liu Z, Zhao Y. Hyaluronic acid-coated shikonin liposomes for the treatment of triple-negative breast cancer via targeting tumor cells and amplification of oxidative stress. *J Drug Deliv Sci Technol.* 2022;70(4):103193. doi: 10.1016/j.jddst.2022.103193
100. Luo K, Yang L, Yan C, *et al.* A dual-targeting liposome enhances triple-negative breast cancer chemoimmunotherapy through inducing immunogenic cell death and inhibiting STAT3 activation. *Small.* 2023;19(40):2302834. doi: 10.1002/sml.202302834
101. Wang T, Yin X, Lu Y, Shan W, Xiong S. Formulation, antileukemia mechanism, pharmacokinetics, and biodistribution of a novel liposomal emodin. *Int J Nanomedicine.* 2012;7:2325-2337. doi: 10.2147/IJN.S31029
102. Chen R, Wang S, Zhang J, Chen M, Wang Y. Aloe-emodin loaded solid lipid nanoparticles: Formulation design and *in vitro* anti-cancer study. *Drug Deliv.* 2015;22(5):666-674. doi: 10.3109/10717544.2014.882446
103. Feng H, Zhu Y, Fu Z, Li D. Preparation, characterization, and *in vivo* study of rhein solid lipid nanoparticles for oral delivery. *Chem Biol Drug Des.* 2017;90(5):867-872. doi: 10.1111/cbdd.13007
104. Sunil Kumar MR, Aithal BK, Udupa N, *et al.* Formulation of plumbagin loaded long circulating pegylated liposomes: *In vivo* evaluation in C57BL/6J mice bearing B16F1 melanoma. *Drug Deliv.* 2011;18(7):511-522. doi: 10.3109/10717544.2011.595840
105. Rachamalla HK, Bhattacharya S, Ahmad A, *et al.* Enriched pharmacokinetic behavior and antitumor efficacy of thymoquinone by liposomal delivery. *Nanomedicine (Lond).* 2021;16(8):641-656. doi: 10.2217/nnm-2020-0470
106. Kontogiannopoulos KN, Assimopoulou AN, Dimas K, Papageorgiou VP. Shikonin-loaded liposomes as a new drug delivery system: Physicochemical characterization and *in vitro* cytotoxicity. *Eur J Lipid Sci Tech.* 2011;113(9):1113-1123. doi: 10.1002/ejlt.201100104
107. Eskandani M, Nazemiyeh H. Self-reporter shikonin-Act-loaded solid lipid nanoparticle: Formulation, physicochemical characterization and geno/cytotoxicity evaluation. *Eur J Pharm Sci.* 2014;59(9):49-57. doi: 10.1016/j.ejps.2014.04.009
108. Li J, Zhao M, Sun M, *et al.* Multifunctional nanoparticles boost cancer immunotherapy based on modulating the immunosuppressive tumor microenvironment. *ACS Appl Mater Interfaces.* 2020;12(45):50734-50747. doi: 10.1021/acsami.0c14909
109. Wu S, Zhu X, Zhang X, *et al.* A thermoresponsive metabolic nanomodulator for achieving photochemotherapy-sensitized cancer immunotherapy. *Chem Eng J.* 2024;499(21):155593. doi: 10.1016/j.cej.2024.155593
110. Li X, Jia X, Niu H. Nanostructured lipid carriers co-delivering lapachone and doxorubicin for overcoming multidrug resistance in breast cancer therapy. *Int J Nanomedicine.* 2018;13:4107-4119. doi: 10.2147/IJN.S163929
111. Xie W, Li Y, Guo Z, *et al.* FePd Nanozyme- and SKN-encapsulated functional lipid nanoparticles for cancer nanotherapy via ROS-boosting necroptosis. *ACS Appl Mater Interfaces.* 2024;16(15):18411-18421. doi: 10.1021/acsami.3c18497
112. Sakpakdeejaroen I, Somani S, Laskar P, Mullin M, Dufes C. Regression of melanoma following intravenous injection of plumbagin entrapped in transferrin-conjugated, lipid-polymer hybrid nanoparticles. *Int J Nanomedicine.* 2021;16:2615-2631. doi: 10.2147/IJN.S293480
113. N'Diaye M, Vergnaud-Gauduchon J, Nicolas V, *et al.* Hybrid lipid polymer nanoparticles for combined chemo- and photodynamic therapy. *Mol Pharm.* 2019;16(9):4045-4058. doi: 10.1021/acs.molpharmaceut.9b00797
114. Yang HZ, Chen JJ, Zhang L, *et al.* A dual responsive nitric oxide/beta-lapachone co-delivery platform for redox imbalance-enhanced tumor therapy. *Eur J Pharm Biopharm.* 2024;201(8):114348. doi: 10.1016/j.ejpb.2024.114348
115. Saeed U, Mahmood R, Fatima B, *et al.* Novel thymoquinone gallate derivative loaded ligand modified quantum dots as pH-sensitive multimodal theragnostic agent for cancer treatment. *Eur J Pharm Biopharm.* 2024;200(7):114312. doi: 10.1016/j.ejpb.2024.114312
116. Jeong SY, Park SJ, Yoon SM, *et al.* Systemic delivery and preclinical evaluation of Au nanoparticle containing beta-lapachone for radiosensitization. *J Control Release.* 2009;139(3):239-245. doi: 10.1016/j.jconrel.2009.07.007
117. Bhattacharjee M, Ghosh A, Das S, *et al.* Systemic codelivery of thymoquinone and doxorubicin by targeted mesoporous silica nanoparticle sensitizes doxorubicin-resistant breast cancer by interfering between the MDR1/P-gp and miR 298 crosstalk. *ACS Biomater Sci Eng.* 2024;10(10):6314-6331. doi: 10.1021/acsbiomaterials.4c01081
118. Zhang Y, Pan H, Yu C, *et al.* Phytoestrogen-derived multifunctional ligands for targeted therapy of breast cancer. *Asian J Pharm Sci.* 2023;18(4):100827. doi: 10.1016/j.ajps.2023.100827
119. Ren S, Song L, Tian Y, *et al.* Emodin-conjugated PEGylation of Fe₃O₄ nanoparticles for FI/MRI dual-modal imaging and therapy in pancreatic cancer. *Int J Nanomedicine.* 2021;16:7463-7478. doi: 10.2147/IJN.S335588
120. Liang J, Tian X, Zhou M, *et al.* Shikonin and chitosan-silver nanoparticles synergize against triple-negative breast cancer through RIPK3-triggered necroptotic immunogenic cell death. *Biomaterials.* 2024;309(6):122608. doi: 10.1016/j.biomaterials.2024.122608
121. Zhao P, Gong L, Chang L, *et al.* Multifunctional Fe-based coordination polymer nano-bomb modified with beta-lapachone and CaO₂ for targeted tumor dual chemodynamic therapy with enhanced ferroptosis and H₂O₂ self-supply. *J Nanobiotechnology.* 2024;22(1):3. doi: 10.1186/s12951-023-02287-2
122. Feng W, Shi W, Liu S, *et al.* Fe(III)-Shikonin supramolecular nanomedicine for combined therapy of tumor via ferroptosis and necroptosis. *Adv Healthc Mater.* 2022;11(2):e2101926. doi: 10.1002/adhm.202101926
123. Feng W, Shi W, Wang Z, *et al.* Enhancing tumor therapy of Fe(III)-shikonin supramolecular nanomedicine via triple ferroptosis amplification. *ACS Appl Mater Inter.* 2022;14(33):37540-37552. doi: 10.1021/acsami.2c11130
124. Zhu X, Zheng L, Zhao P, *et al.* Fe/Cu bimetallic nanozyme Co-assembled with ¹⁷⁷Lu and tanshinone for quadruple-synergistic tumor-specific therapy. *Adv Healthc Mater.* 2024;14:e2402696. doi: 10.1002/adhm.202402696
125. Shi W, Feng W, Li S, *et al.* Ferroptosis and necroptosis produced autologous tumor cell lysates Co-delivering with combined immunoadjuvants as personalized *in situ* nanovaccines for antitumor immunity. *ACS Nano.* 2023;17(15):14475-14493. doi: 10.1021/acs.nano.3c00901
126. Asrorov AM, Gu Z, Li F, Liu L, Huang Y. Biomimetic camouflage delivery strategies for cancer therapy. *Nanoscale.* 2021;13(19):8693-8706. doi: 10.1039/d1nr01127h
127. Wu M, Ling W, Wei J, *et al.* Biomimetic photosensitizer nanocrystals trigger enhanced ferroptosis for improving cancer treatment. *J Control*

- Release*. 2022;352(12):1116-1133.
doi: 10.1016/j.jconrel.2022.11.026
128. Zhang W, Li L, Wu Y, et al. Biomimetic iron-based nanoparticles remodel immunosuppressive tumor microenvironment for metabolic immunotherapy. *Int J Nanomedicine*. 2024;19:9333-9349.
doi: 10.2147/IJN.S473463
129. Liu W, Wu J, Ji X, et al. Advanced biomimetic nanoreactor for specifically killing tumor cells through multi-enzyme cascade. *Theranostics*. 2020;10(14):6245-6260.
doi: 10.7150/thno.45456
130. Wang H, Tang Y, Fang Y, et al. Reprogramming tumor immune microenvironment (TIME) and metabolism via biomimetic targeting codelivery of shikonin/JQ1. *Nano Lett*. 2019;19(5):2935-2944.
doi: 10.1021/acs.nanolett.9b00021
131. Cui J, Wang X, Li J, et al. Immune exosomes loading self-assembled nanomicelles traverse the blood-brain barrier for chemo-immunotherapy against glioblastoma. *ACS Nano*. 2023;17(2):1464-1484.
doi: 10.1021/acsnano.2c10219
132. Han S, Bi S, Guo T, et al. Nano co-delivery of plumbagin and dihydrotanshinone i reverses immunosuppressive TME of liver cancer. *J Control Release*. 2022;348(8):250-263.
doi: 10.1016/j.jconrel.2022.05.057
133. Soni P, Kaur J, Tikoo K. Dual drug-loaded paclitaxel-thymoquinone nanoparticles for effective breast cancer therapy. *J Nanopart Res*. 2015;17(1):1-12.
doi: 10.1007/s11051-014-2821-4
134. Li H, Tong Y, Bai L, et al. Lactoferrin functionalized PEG-PLGA nanoparticles of shikonin for brain targeting therapy of glioma. *Int J Biol Macromol*. 2018;107(2):204-211.
doi: 10.1016/j.ijbiomac.2017.08.155
135. Peng J, Zhou W, Xia X, et al. Encapsulation of acetylshikonin by polyamidoamine dendrimers for preparing prominent nanoparticles. *AAPS PharmSciTech*. 2014;15(2):425-433.
doi: 10.1208/s12249-014-0074-2
136. Jiang X, Ma M, Li M, et al. Preparation and evaluation of novel emodin-loaded stearic acid-g-chitosan oligosaccharide nanomicelles. *Nanoscale Res Lett*. 2020;15(1):93.
doi: 10.1186/s11671-020-03304-1
137. Wang X, Guo Y, Qiu L, et al. Preparation and evaluation of carboxymethyl chitosan-rhein polymeric micelles with synergistic antitumor effect for oral delivery of paclitaxel. *Carbohydr Polym*. 2019;206(4):121-131.
doi: 10.1016/j.carbpol.2018.10.096
138. Huang Y, Lu J, Gao X, et al. PEG-derivatized embelin as a dual functional carrier for the delivery of paclitaxel. *Bioconjug Chem*. 2012;23(7):1443-1451.
doi: 10.1021/bc3000468

Received: December 31, 2024

Revised: March 12, 2025

Accepted: March 24, 2025

Available online: May 8, 2025